

II.K.15 Hydrogenases of Methanococcus Maripaludis

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Objectives

The research examines the enzymes and pathways in H_2 metabolism in methanogens, especially as they relate to energy conservation. In addition, the metabolism of formate, from which H_2 is produced, is investigated.

Technical Barriers

The research should lead to an enhanced understanding of how two universal forms of biological energy, the chemical energy contained in the bonds of molecular hydrogen and the energy stored in ion gradients that form across cell membranes, can be interconverted.

Abstract

The methanogens catalyze a major component of the Earth's H_2 cycle. They are especially active in anaerobic environments where they are the primary consumers of fermentatively produced H_2 . These strictly anaerobic Archaea have evolved unique adaptations to H_2 metabolism, many of which are poorly understood. Our research examines the enzymes and pathways in H_2 metabolism in methanogens, especially as they relate to energy conservation (Figure 1). In addition, the metabolism of formate, from which H_2 is produced, is being investigated. The model species of choice is the hydrogenotroph *Methanococcus maripaludis*. Transcriptome and proteome studies have determined the regulatory effects of H_2 limitation. Studies of protein complexes indicate that the heterodisulfide reductase is membrane-associated and also forms a complex with certain hydrogenases and with formate dehydrogenase.

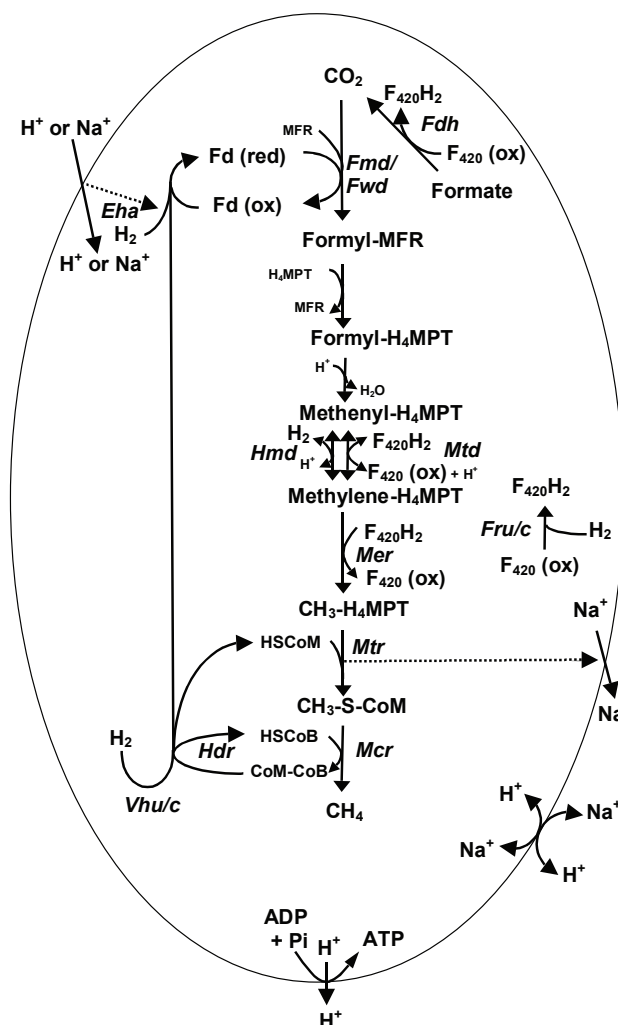


FIGURE 1. Methanogenic pathway of the hydrogenotrophic methanogens. Abbreviations for enzymes are in italics: *Eha*, energy-conserving hydrogenase A; *Fdh*, formate dehydrogenase; *Fmd/Fwd*, formyl-methanofuran dehydrogenase; *Fru/c*, F_{420} -reducing hydrogenase; *Hdr*, heterodisulfide reductase; *Hmd*, H_2 -dependent methylene- H_4 -methanopterin dehydrogenase; *Mcr*, methyl-coenzyme M reductase; *Mer*, methylene- H_4 -methanopterin reductase; *Mtd*, F_{420} -dependent methylene- H_4 -methanopterin dehydrogenase; *Mtr*, methyl- H_4 -methanopterin-coenzyme M methyltransferase; *Vhu/c*, F_{420} -nonreducing hydrogenase. Dotted arrows indicate coupling of metabolic steps with membrane ion gradients.

The analysis of mutants has suggested that two alternative pathways function in the interconversion of reduced coenzyme F_{420} and H_2 . Finally, growth yield studies with chemostats suggest that two alternative pathways of energy conservation function under differing conditions of H_2 availability.

Progress report

Function of the energy-conserving hydrogenase Ehb and global regulation in an Ehb mutant. We constructed a mutant of *Methanococcus maripaludis* lacking the energy-conserving hydrogenase Ehb and obtained evidence that the mutant was deficient in biosynthetic CO₂ assimilation. Microarrays and differential proteomics confirmed that the mRNA and protein levels for the enzymes of CO₂ assimilation were elevated in the mutant. These results were consistent with the hypothesis that Ehb coupled the reduction of a low potential ferredoxin with a chemiosmotic membrane gradient. This ferredoxin would then be used to drive the highly endergonic reactions of acetyl-CoA synthase, pyruvate oxidoreductase, and other biosynthetic enzymes that assimilate CO₂.

Identification of genes regulated at the mRNA level by H₂ limitation. Regulation by H₂ availability in methanogens is important because it can identify cellular components that play key roles in H₂ utilization, electron flow, and energy conservation. We devised a system for growth of *M. maripaludis* in chemostats, where growth rate can be controlled and specific nutrient limitations can be imposed. By comparing H₂ limitation to two other nutrient limitations (phosphate limitation and leucine limitation), we were able to discern which genes were regulated by each condition. In addition, we varied growth rate without changing the limiting nutrient, and were able to determine which genes were regulated by growth rate. H₂ limitation markedly increased the mRNA abundance for genes encoding reactions that involve F₄₂₀ as electron acceptor or electron donor. Thus, mRNA levels for *fru*, *mtd*, *mer*, and *fdh* were markedly higher in H₂-limited cultures.

Development of a membrane vesicle system and characterization of the particulate heterodisulfide reductase (pHdr). Inverted membrane vesicles have been indispensable tools in the analysis of chemiosmotic energy coupling, including studies of *Methanosarcina*. We prepared inverted vesicles of *M. maripaludis* by methods developed for *Methanosarcina* strains. The vesicles were assayed for heterodisulfide reductase (Hdr) activity. Prior to these studies, the Hdr from hydrogenotrophic methanogens was reported to be soluble. Because this enzyme was believed to play a fundamental role in coupling methanogenesis to the proton motive force, it was expected to be membrane bound. To examine this point more fully, an assay was developed for the Hdr. For extracts prepared by the French press, 34% of the activity was associated with the membrane fraction. Moreover, the specific activity of washed membranes was 38 nmol min⁻¹ mg⁻¹, or comparable to the H₂- or coenzyme F₄₂₀-dependent activity observed by others. The specific activity of the membranes was also higher than that of the soluble fraction, suggesting enrichment of the Hdr. These results

indicate that Hdr activity is membrane-associated and has implications for our model of energy conservation.

Co-purification of possible energy-conserving components with heterodisulfide reductase. We have begun experiments to use the Hdr to “pull down” components of a complex that may be involved in energy conservation. We constructed strains expressing oligo-his tagged HdrA_c and HdrB1, two subunits of heterodisulfide reductases, and conducted anaerobic nickel affinity purifications of cell extracts. We then separated purified proteins by 2-D gel electrophoresis and identified the spots by MALDI-TOF mass spectrometry. Co-purifying proteins included the VhuD subunit of non-F₄₂₀ reducing hydrogenase and the FdhA and FdhB subunits of formate dehydrogenase, as well as additional subunits of Hdr. Note that Fdh is expressed in high levels even in H₂-grown cells. These results are exciting because they suggest that all three enzymes are part of a complex. The complex of Hdr with non-F₄₂₀ reducing hydrogenase has been observed before and is consistent with direct transfer of electrons from the hydrogenase to Hdr. The complex of Hdr with Fdh is novel and suggests that there may be a direct transfer of electrons from formate to Hdr.

Analysis of membrane proteins by Blue Native PAGE and mass spectrometry. Protein complexes in the membrane fractions of *M. maripaludis* were analyzed using Blue Native PAGE (BN-PAGE) and mass spectrometry. About one-half of the total hydrogenase activity was associated with membrane preparations. Upon BN-PAGE, 16 proteins were identified by in-gel digestion and MALDI-FTICR. Highly expressed cytoplasmic proteins, such as ribosomal and S-layer proteins as well as subunits of the methyl coenzyme M reductase were not detected, indicating efficient separation of the membranes from soluble proteins. The identified proteins included components of three well-known membrane protein complexes, A₁A₀ ATPase, glutamate synthase, and methyl-H₄MPT-HSCoM methyltransferase (Mtr). An additional band contained components of the proteasome. Presumably, this very high molecular weight complex copurified with the membranes. Two bands possessed high levels of hydrogenase activity. Mass spectrometry also identified subunits of the coenzyme F₄₂₀-reducing hydrogenase (Fru) and energy conserving hydrogenase B (Ehb). The large subunit of coenzyme F₄₂₀-nonreducing hydrogenase (Vhu) was also identified. Other enzymes co-migrated with these hydrogenases, suggesting that they may form physiologically important complexes. Subunits of the heterodisulfide reductase and formylmethanofuran dehydrogenase co-migrated with the F₄₂₀-nonreducing hydrogenase Vhu. Similarly, a complex of Vhu and heterodisulfide reductase has also been detected by BN-PAGE of *Methanothermobacter thermoautotrophicus*. The formation of a complex of these three enzymes

is consistent with models that couple heterodisulfide reduction to the initial step of CO₂ reduction and the existence of contiguous genes encoding of these proteins. However, this complex is predicted to be soluble and not membrane-bound, as observed here. One band contained the Fru with more components of the formyl-MF dehydrogenase. This seems to provide presumptive evidence for multiple complexes between the formyl-MF dehydrogenase and hydrogenases.

Mutants in F₄₂₀-reducing hydrogenases (Fru/Frc), F₄₂₀-dependent methylenetetrahydromethanopterin dehydrogenase (Mtd), and H₂-dependent methylenetetrahydromethanopterin dehydrogenase (Hmd). One approach we have taken is to make mutants deficient in the hydrogenases and in relevant steps in methanogenesis. We successfully made a strain containing in-frame deletions in the alpha subunits of both F₄₂₀-reducing hydrogenases, $\Delta fruA-\Delta frcA$. This mutant grew well on both H₂ and formate. Because F₄₂₀H₂ is required for methanogenesis, this finding suggests that there is an alternate pathway for the reduction of F₄₂₀ from H₂. That pathway could be the previously proposed Hmd-Mtd cycle, in which Mtd working in reverse reduces F₄₂₀ (Fig. 1). In addition, we found that during growth on formate, H₂ is produced in the headspace during growth and methanogenesis. The product of formate oxidation is F₄₂₀H₂, and H₂ production evidently occurs from F₄₂₀H₂. There was no decrease in H₂ accumulation in the $\Delta fru-\Delta frc$ mutant, suggesting that the Hmd-Mtd cycle can also function to produce H₂. We have also made an *mtd* mutant (Δmtd) and an *hmd* mutant (Δhmd). The only growth defect for either mutant was a lag before growth of the Δmtd mutant on formate. During this lag, the Δmtd mutant accumulated H₂ in the headspace to a higher level than did the other strains. These results are consistent with a complex interaction between Fru/c, Mtd and Hmd. During growth on formate, H₂ is produced by either the Fru/c or Mtd-Hmd systems, presumably in equilibrium with the formate concentration. However, in the Δmtd mutant, methanogenesis is blocked at the methenyl-H₄MPT step at low levels of H₂. Only when H₂ accumulates to higher levels can the low affinity Hmd become active and methanogenesis continue.

We studied the production of H₂ from formate in resting cells. The characteristics of this process may have implications for the interactions of redox enzymes and whether H₂ is an obligatory intermediate during growth on formate. Deletion of the F₄₂₀-reducing hydrogenase, Fru, severely decreased H₂ production, suggesting that the major pathway of H₂ production comprised formate dehydrogenase and Fru. Because the Δfru mutant retained 20% of the wild type H₂ production activity, an additional pathway was present. Deletion of the *mtd* or *hmd*, decreased the activity by 20% of the wild-type level, indicating that the second pathway was comprised of formate dehydrogenase-Mtd-Hmd.

High rates of formate-dependent H₂ production were consistent with the hypothesis that the H₂ utilization reactions of methanogenesis were reversible and demonstrated the potential of *M. maripaludis* for the microbial production of H₂ from formate.

Growth yields under hydrogen limited chemostat cultures—the uncoupling phenomenon.

Hydrogenotrophic methanogens are known to “uncouple” growth from methanogenesis under some circumstances, that is, the cell yield per mol of methane produced or Y_{CH₄} is less under some conditions. We used our chemostat system to determine conditions that affect cell yield of *M. maripaludis*. A H₂-limited chemostat produced three to four-fold less methane than a phosphate-limited chemostat. Growth yields (Y_{CH₄}, g dry wt of cells per mole CH₄) were 3.18 under H₂ limitation and 0.94 under phosphate limitation. Hence, uncoupling of methanogenesis from growth occurs under phosphate-limited conditions or when H₂ is in excess. This observation indicates that the pathway of electron flow and/or energy coupling varies depending upon the growth conditions. Thauer’s recent findings regarding electron flow between the heterodisulfide reductase complex and the formyl-MFR dehydrogenase suggests a biochemical mechanism for the uncoupling. In methanococci, electrons for formyl-MFR dehydrogenase could come predominately from the heterodisulfide reductase complex under H₂-limited conditions, conserving energy, but come predominately from the Eha hydrogenase under H₂ excess, depleting the membrane potential (Figure 1). While the latter pathway would impose a thermodynamic disadvantage, it could provide a kinetic advantage.

Future directions

We will test the model for energy coupling that is outlined in Fig. 1 above. The coupling of the exergonic heterodisulfide reductase and the endergonic formyl-MFR dehydrogenase reactions are pivotal. We propose that two mechanisms function: the low potential electron donor for formyl-MFR dehydrogenase is generated either by the Eha hydrogenase in a membrane-dependent reaction or by coupling to the Vhu/c-Hdr complex. In addition, we will test a model for electron flow during growth on formate, and determine whether H₂ is a required intermediate.

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