Maximizing Photosynthetic Efficiencies and Hydrogen Production in Microalgal Cultures

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Project ID # PD15

This presentation does not contain any proprietary or confidential information

Overview

Timeline

- Start: October 2000
- End: December 2004
- Completion: 40%

Budget

- Funding in FY04
- DOE: \$200 k, UCB: \$50 k
- Funding for FY05
 DOE: \$200 k, UCB: \$75 k

Barriers addressed

 Low Light Utilization Efficiency in Photobiological Hydrogen Production due to a Large Photosystem Chlorophyll Antenna Size (Barrier X).

Partners

• NREL, ORNL, DaimlerChrysler

Objectives and Approach

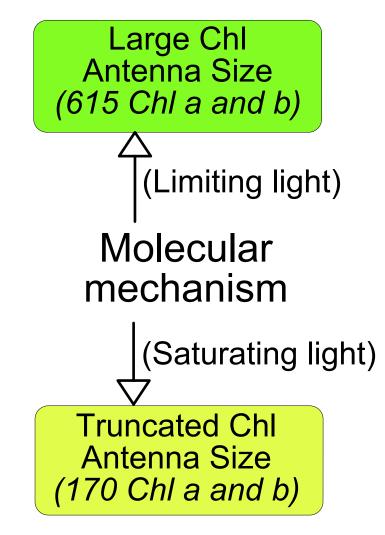
Objective: Minimize the chlorophyll antenna size of photosynthesis to maximize solar conversion efficiency in green algae.

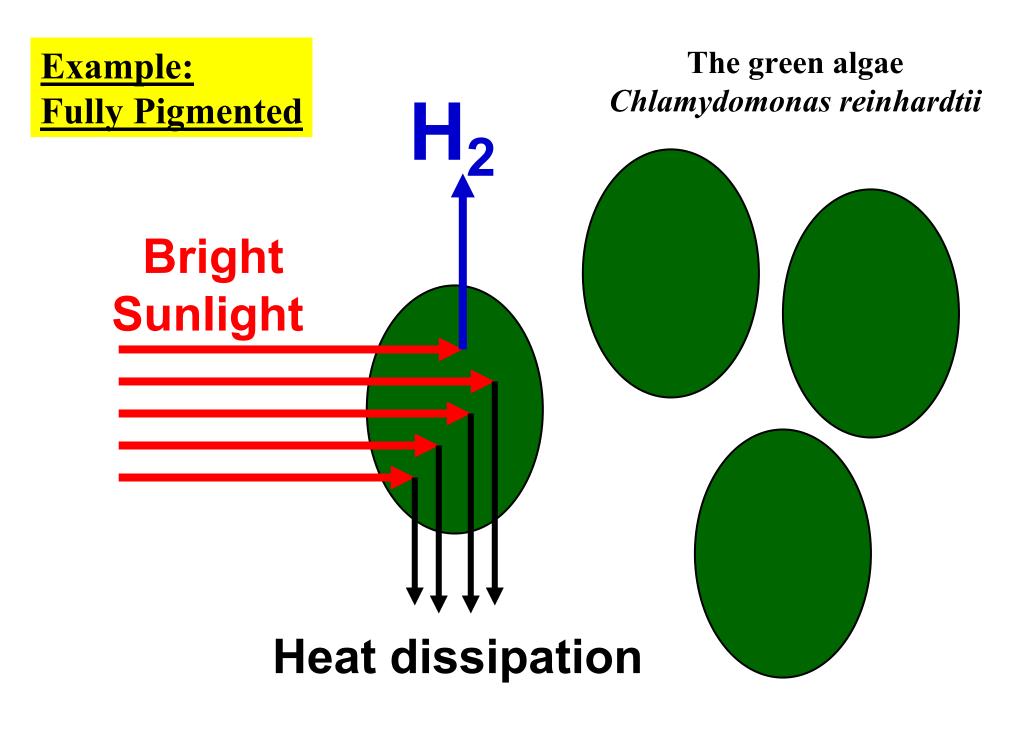
(Identify and characterize genes that regulate the ChI antenna size in the model green alga *ChIamydomonas reinhardtii.* Apply these genes to other green algae, as needed.)

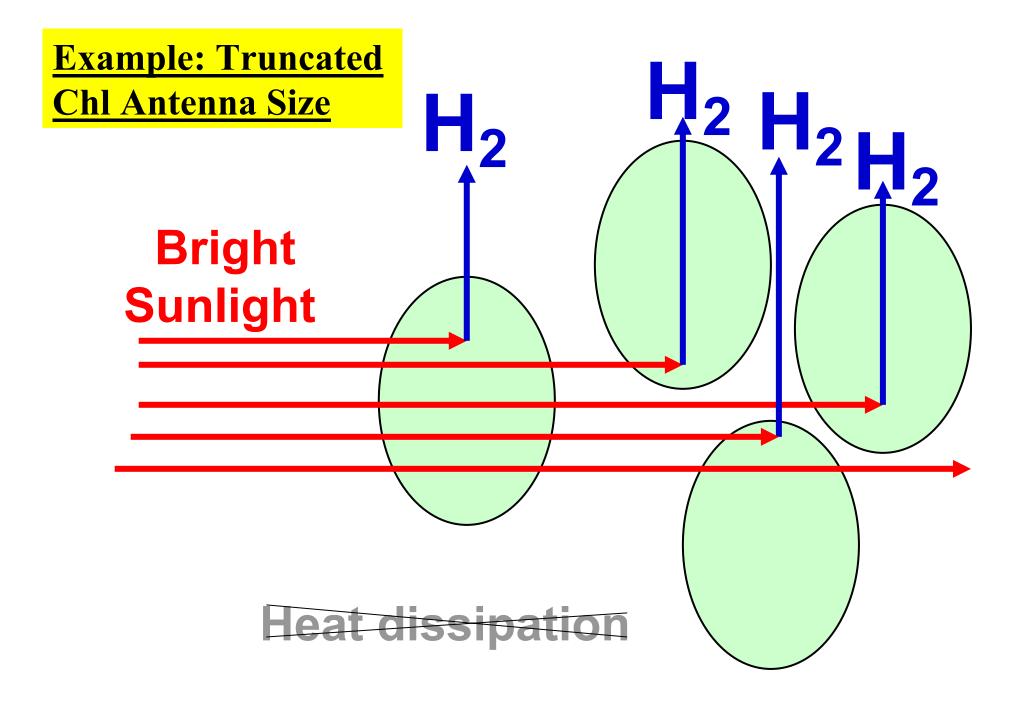
<u>Approach</u>: Interfere with the molecular mechanism for the regulation of the chlorophyll antenna size.

(Employ DNA insertional mutagenesis and highthroughput screening to isolate tagged green algae with a smaller ChI antenna size.)

Regulation of the Chl antenna size

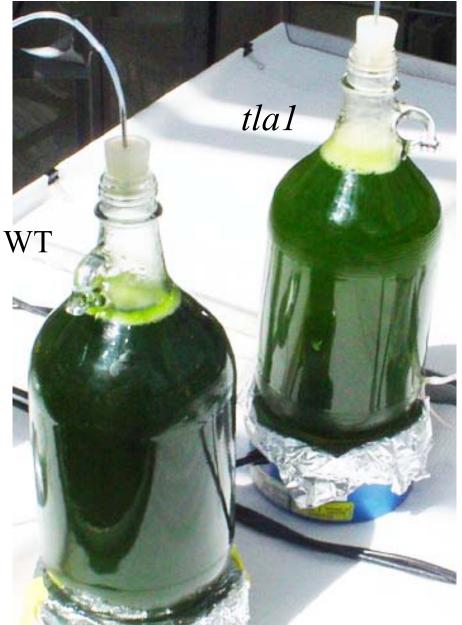






Measurement in Scale-up Cultures

Cultures in the Greenhouse



Parameter	<u>WT</u>	<u>tla1</u>					
Cell/mL (x10 ⁶)	6.36	10.0					
[Chl] (uM)	25.6	15.4					

Benefits from this Project

Truncating the Chlorophyll antenna size of microalgae would benefit photobiological:

H₂ production,

- carbon sequestration,
- biomass accumulation,
- waste water treatment,
- other bio-fuels generation.

Technical Barriers and Targets

- <u>Barrier X</u>: Low Light Utilization Efficiency in Photobiological Hydrogen Production due to a Large Photosystem Chlorophyll Antenna Size.
- Light Utilization Efficiency of WT green algae: 3-5%
- <u>Theoretical maximum efficiency</u>: ~30%
- <u>Target for 2010</u>: Reach a 15% Utilization Efficiency of Absorbed Light Energy.

Chl Antenna Size vs Light Utilization Efficiency Utilization Efficiency of Absorbed Light Energy Achievement in 2004: 15%

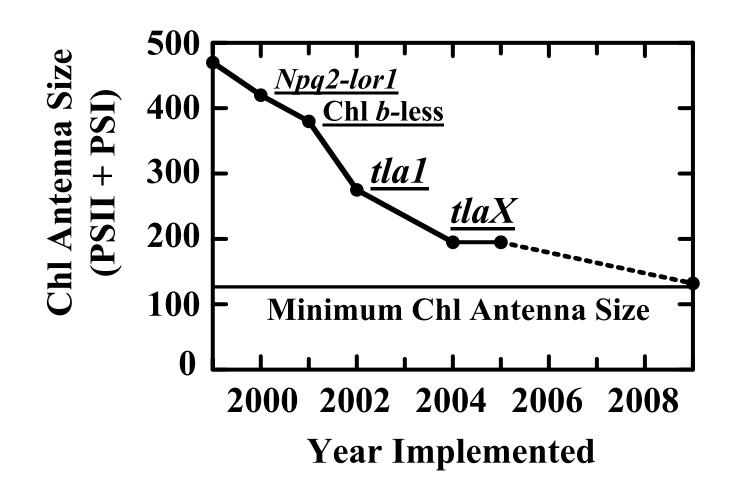
- Wild type antenna size = <u>470 Chl molecules</u> (100%) (PSII=230; PSI=240) Photon use efficiency of WT photosynthesis = ~6-10% <u>Utilization Efficiency of Absorbed Light Energy by WT: ~3-5%</u>
- *tla1* antenna size = <u>275 Chl molecules</u> (59% of control) (PSII=115; PSI=160) Photon use efficiency of *tla1* photosynthesis = ~20% <u>Utilization Efficiency of Absorbed Light Energy by *tla1*: ~10%</u>

2004 Year Accomplishment

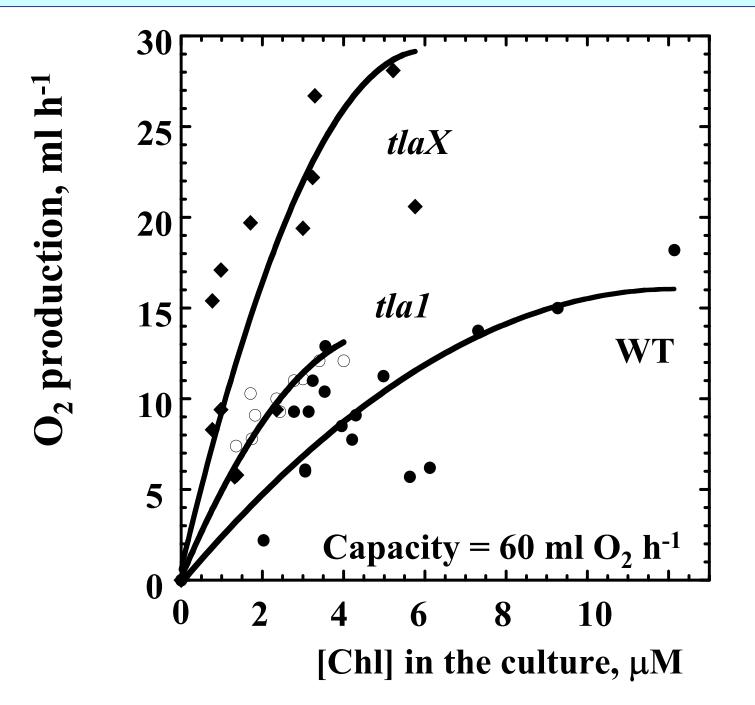
- tlaX antenna size = <u>195 Chl molecules</u> (42% of control) (PSII=80; PSI=115) Photon use efficiency of *tlaX* photosynthesis = ~30% <u>Utilization Efficiency of Absorbed Light Energy by *tlaX*: ~15%</u>
- Long-term goal: 132 Chl molecules (28% of control) (PSII=37; PSI=95)
 Photon use efficiency of photosynthesis goal = ~60%
 <u>Utilization Efficiency of Absorbed Light Energy goal: ~30%</u>

Project Timeline

Chlorophyll Antenna Size in Chlamydomonas



Productivity in Scale-up Cultures



Current Technical Accomplishments Analysis of the *tla1* and *tlaX* mutants

• Molecular analysis of the *tla1* mutation.

DNA insertion site in the *tla1* mutant has been mapped.Genomic, cDNA and protein sequences for the *Tla1* gene are at hand.Complementation of the *tla1* mutant with the *Tla1* gene completed,Analysis of the complemented strains in progress.

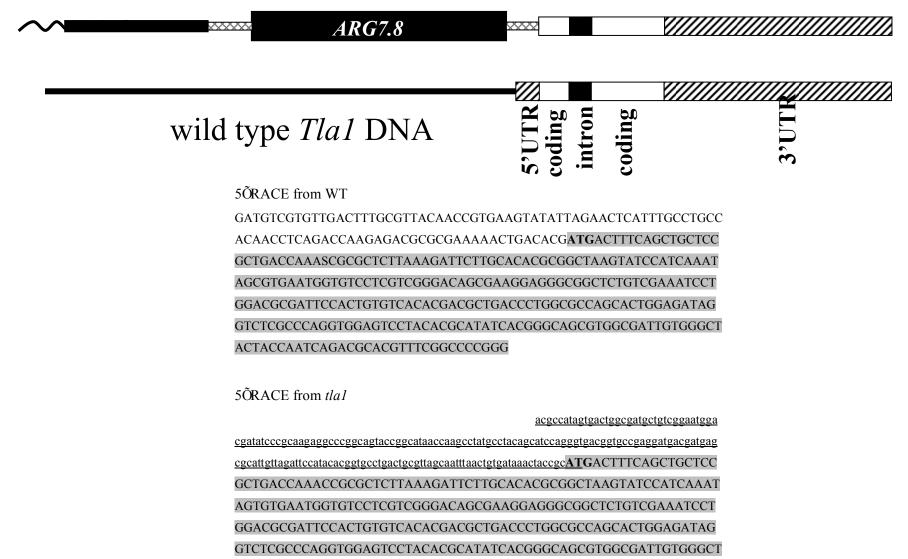
Biochemical analysis of the *tla1* mutation.

Antibodies against the Tla1 protein are being raised. Hydropathy plot of the Tla1 protein measured. Sequence homologies for the Tla1 protein completed.

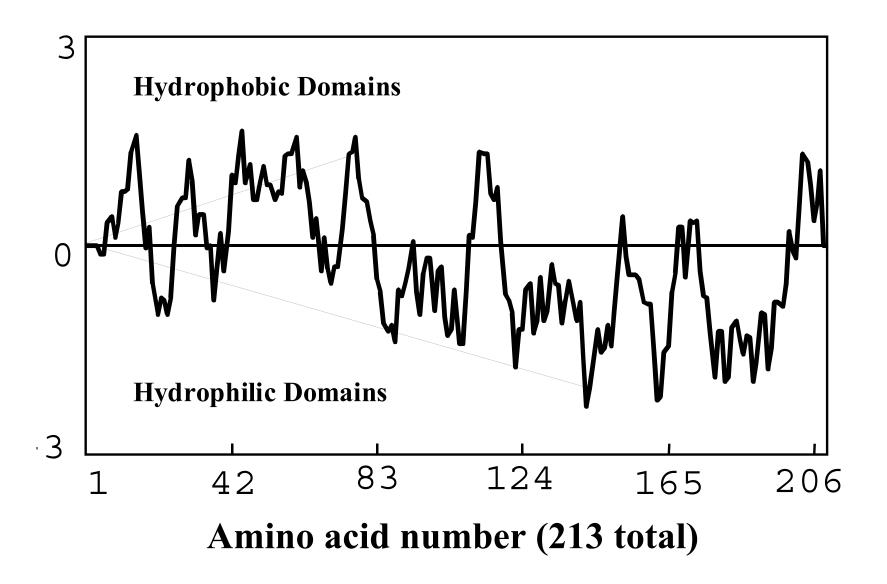
- Functional analysis of the *Tla1* gene. Regulation of the chlorophyll antenna size by the *Tla1* gene.
- **Biophysical and biochemical analyses of the** *tlaX* **mutant.** Chlorophyll antenna size, relative productivity, LHC expression levels.

Mapping of the *tla1* mutation and WT *Tla1* gene structure

tla1 mutant DNA



Hydropathy plot of the Tla1 deduced amino acid sequence



Sequence homologies for the Tla1 protein

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О.	sativa	MG-	-AE(CKYE	EVAQ	V <mark>AY\</mark>	/KLA	LHAI	KHE	2AAA	VN	GLI	JVG	RLL	DGA	ASI	PAAV	7 <mark>VS</mark>	IADZ	77
H.	sapiens	MG-		-EVI	EISA	L <mark>AY</mark>	/KMC	LHAA	RYE	PHAA	VN	GLF	'LA	PAP	R		SGE	G <mark>LC</mark>	LTDO	27
D.	melanogaster	MC-		-DYF	KVSE	R <mark>AY</mark>	KLI	FHAA	KYE	PHQA	VN	GLI	'I'Y	EKT	s	I	KGSÇ	O <mark>VE</mark>	IVDA	4 :
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	reinhardtii	CHT				-	-						-							
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D.	melanogaster	FHQ	CLY	VTPN	IAEV	ALMI	IDA	HAER	EG-	-LVI	AG	YYA	AP	ENF	YDN	IQVI	OKTI	PAA	KIAI)]
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	thaliana	RYFPQ																		
-	sativa	RNFPR																		
	sapiens	EFFPD																		
D.	melanogaster	ENFKN														-SI				
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C.	reinhardtii	LREEF	FVMI	FKOI	KH <mark>R'</mark>	TLH	FEE	HLDD	AGK		NK	GFA	SS	v-ĸ	FLI	•P	0	;NA	L	
А.	thaliana	VLSDY																		
	sativa	VLADH																		
	sapiens	MVGAL																		
	-				-					-										
D_{\cdot}	melanogaster	EGVSL	LTKI	RG	-AM <mark>R</mark> I	DLVI	FDN	HLDN	IPDK	NWT	'ND	FLN	IOP	LND	LOK	LY-			-	

Functional and regulation analysis of the *Tla1* gene

Regulation of the chlorophyll antenna size by the Tla1 gene

- Expression levels of the *Tla1* gene increase with the level of irradiance.
 (leads to smaller Chl antenna size)
- In the *tla1* mutant, levels of *Tla1* gene transcripts are higher than in the WT.
 (leads to smaller Chl antenna size)
- When expression level of the *Tla1* gene is high, expression levels of the *Lhcb* and *Cao* genes are low and Chl-protein content is also low.

(leads to smaller Chl antenna size)

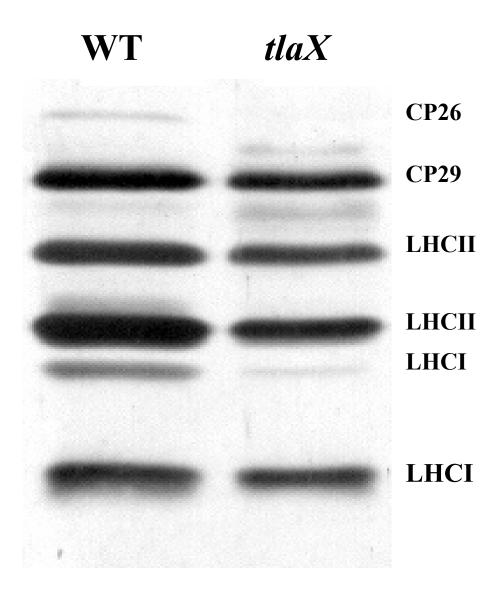
Tentative conclusion

• Enhancing the expression level of the *Tla1* gene minimizes the Chl antenna size of photosynthesis.

Application Hypothesis

• Over-expression of the ubiquitous *Tla1* gene would Maximize Photosynthetic Efficiencies and Hydrogen Production in Microalgal Cultures.

Biochemical analysis of the Chl antenna proteins



Responses to Previous Year Reviewers Comments

<u>Systems analysis and engineering should be included</u>

This is being done in concert with the other two Photobiological Hydrogen production projects at NREL and ORNL (see <u>http://www.nrel.gov/docs/fy04osti/35593.pdf</u>).

• PI should play team leadership role

Devised the concept and helped draft the experimental roadmap of the "**Integrated Biological Hydrogen Production**" multiyear plan for the DOE HFCIT program.

<u>Combine the four separate mutations as highest priority</u>

This is being planned, requires a full-time geneticist who would perform the crosses and successfully analyze the offspring (a difficult position to fill).

<u>Consider second row of mutations and screen on best mutant "tlaX"</u>

Although contemplated, the high throughput screening of *tlaX* mutants for strains with an even smaller Chl antenna size may be difficult to do. It may be more practical to mutagenize and screen wild type green algae and then, to combine the genetic properties of *tla*-type mutants.

<u>Need to more closely relate alga types (mutant vs wild type) in H₂ as well as O₂ production </u>

Productivity measurements on the basis of biomass and O_2 are routinely conducted at Berkeley in wild type and each of the *tla* mutants. The sulfur-deprivation method is not suitable for the measurement of H₂-production by the *tla* mutants. Until an alternative steady-state method of green algal H₂-production can become available, only the ORNL "continuous sparging with He" method is suitable for testing the *tla* mutants.

Future Work

Remainder of FY 2005

1. Complete the biochemical, molecular and genetic analyses of the *tla1 and tlaX* strains. Publish *tla1-* and *tlaX*-related findings.

FY 2006 and Beyond

- 1. Functionally characterize the corresponding *tla1* and *tlaX* genes (how do they work?)
- 2. Establish transformation (sense and antisense) protocols with *Tla*-type genes to enhance the downregulation of the ChI antenna size in *Chlamydomonas reinhardtii.*
- 3. Perform genetic crosses to combine different *tla*-type properties.

MORE ON POSTER PDP29

Publications and Presentations

- Posewitz MC, Smolinski SL, Kanakagiri S, Melis A, Seibert M, Ghirardi ML (2004) Hydrogen photo-production is attenuated by disruption of an isoamylase gene in *Chlamydomonas reinhardtii.* Plant Cell 16: 2151-2163
- Komine Y and Melis A (2004) *Chlamydomonas reinhardtii* thioredoxin-like protein mRNA, complete cds; nuclear gene. GenBank Accession Number AY762116
- Melis A, Seibert M and Happe T (2004) Genomics of green algal hydrogen research. Photosynth. Res. 82: 277-288
- Melis A (2005) Bioengineering of green algae to enhance photosynthesis and hydrogen production. Chapter 12 in *Artificial Photosynthesis: From Basic Biology to Industrial Application,* AF Collins and C Critchley (eds.), Wiley-Verlag & Co., In Press
- White A and Melis A (2004) Maximizing Light Utilization Efficiency and Hydrogen Production in Microalgal Cultures. Abstracts of the 13th Western Photosynthesis Conference, Asilomar Conference Center, Pacific Grove, CA. January 8-11, p. 33.
- Melis A, Polle J and Kanakagiri S (2004) Genes for the regulation of the light-harvesting chlorophyll antenna size in *Chlamydomonas reinhardtii*. Plant Biology 2004, Abstract # 219 in the American Society of Plant Biologists 2004 Annual Meeting Program Book. p. 80.
- Melis A, Polle JE and Kanakagiri S (2004) Genes for the regulation of the light-harvesting chlorophyll antenna size in *Chlamydomonas reinhardtii*. Abstract # 259 of the 13th International Congress of Photosynthesis. pp. 112-113

Hydrogen Safety

The most significant hydrogen hazard associated with this project is:

The presence of pressurized cylinders with hydrogen, nitrogen and argon that are employed in the conduct of this work

These are safely anchored in appropriately designed berth spaces.

Hydrogen Safety

Our approach to deal with this hazard is:

Training of personnel in general, and specific aspects of safety for this project in particular, is mandatory for all employees in this department.

The small amounts (ml quantities) of H₂ involved in this work do not entail a significant hazard, nor do they pose an accident scenario.

Safety oversight is maintained by the University's Environmental Health and Safety office (EH&S).