



# **Bioinspired Composite Nanomaterials for Photocatalytic Hydrogen Production**

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Center for Bioinspired Nanomaterials  
Pleotint LLC

DOE Project ID#: PDP36



# Overview



## Timeline

- Start - Oct. 2005
- End - Sept. 2009

## Budget

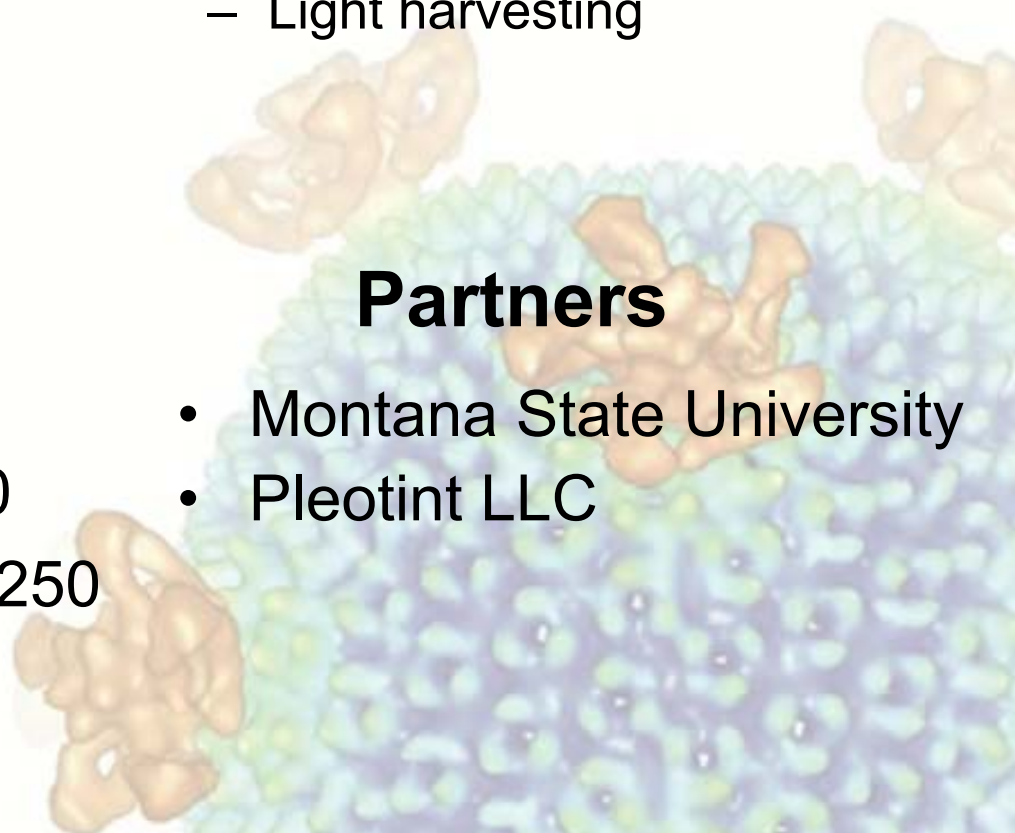
- Total project funding
  - \$1,491,250
  - DOE \$1,193,000
  - Contractor \$298,250

## • Barriers addressed

- Enzyme stability/durability
- Oxygen sensitivity
- Light harvesting

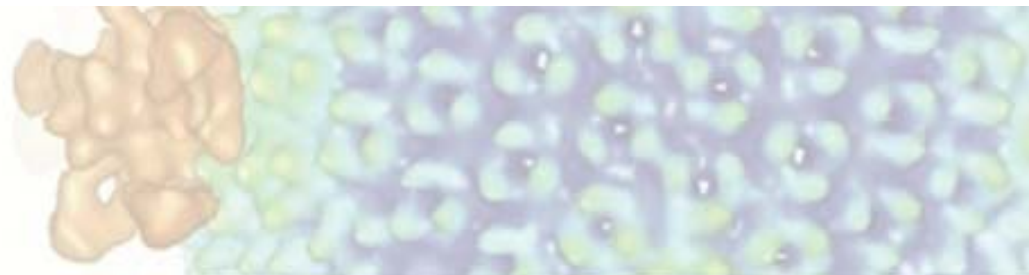
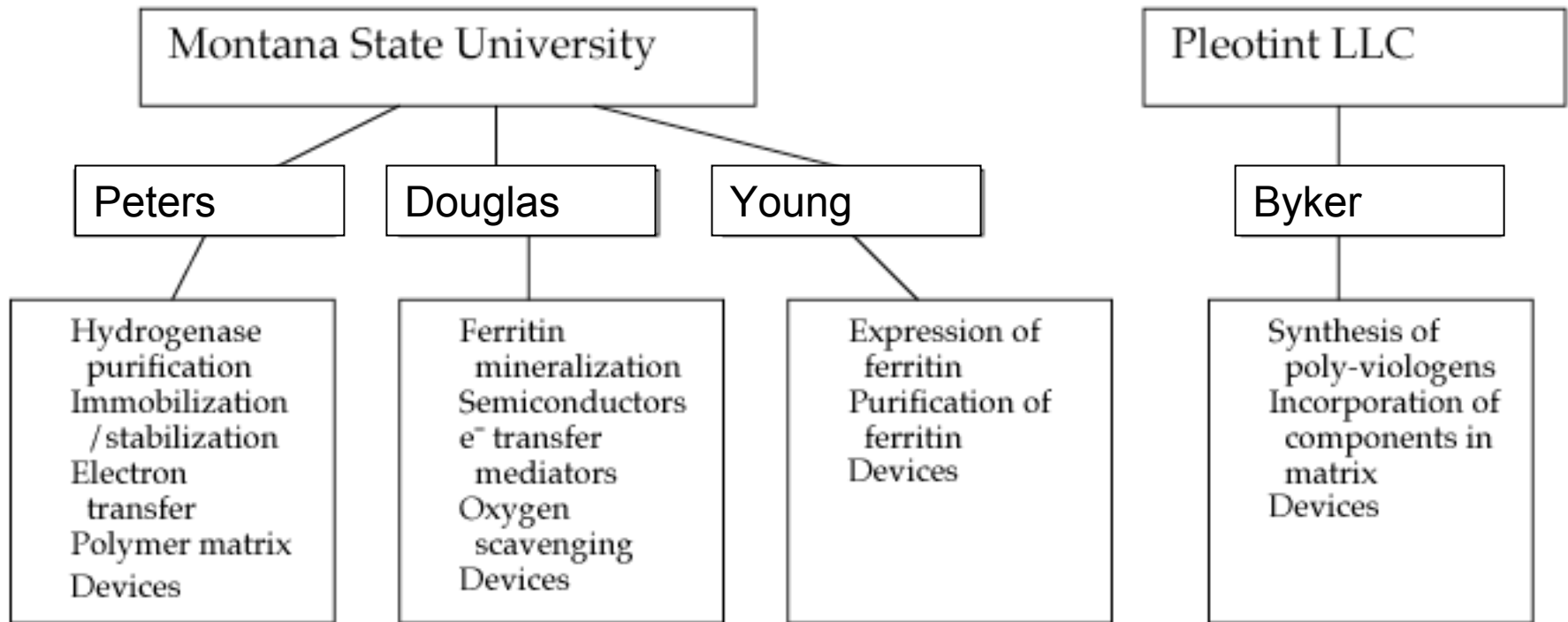
## Partners

- Montana State University
- Pleotint LLC





# Overall Project Structure





# Objectives



1. Optimize the hydrogenase stability and electron transfer
2. Optimize the semiconductor nano-particle photocatalysis, oxygen scavenging, and electron transfer properties of protein nano-cages
3. Gel/Matrix immobilization and composite formulation of nano-materials and hydrogenase
4. Device fabrication for H<sub>2</sub> production

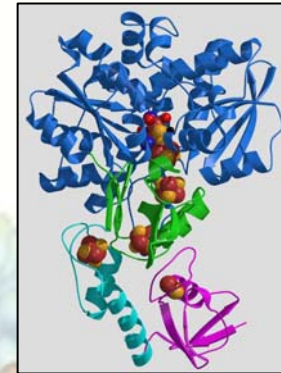


# Approaches

## Couple Different Catalyst Systems for Light Driven Hydrogen Generation

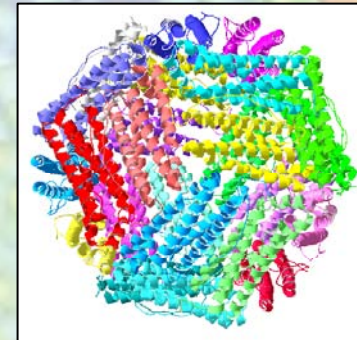
Biological catalysts (Hydrogenases)

- stabilization/immobilization
- electron transfer



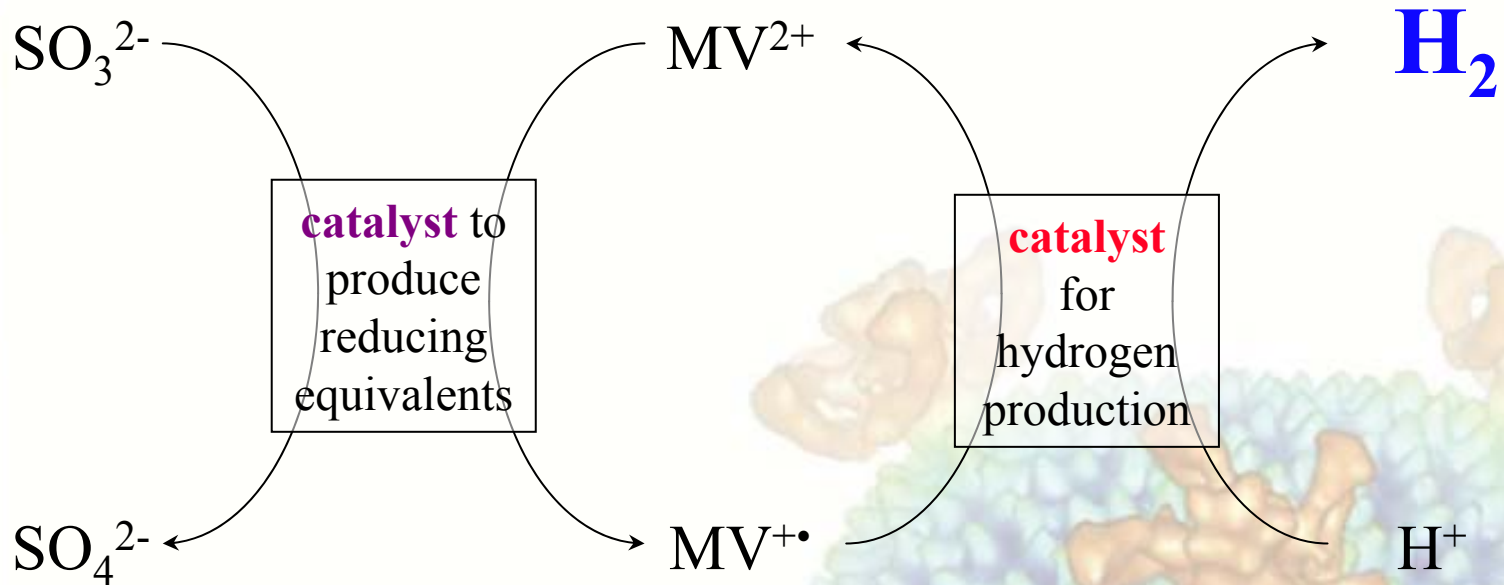
Nanoparticle Photocatalysts

- light harvesting
- O<sub>2</sub> scavenging

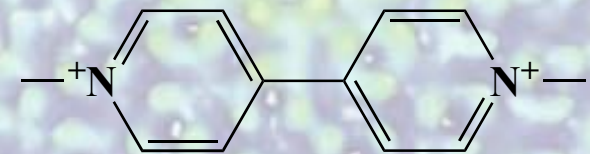




# Hydrogen from Water: Coupled Reactions



**GOAL:** use a **catalyst** to reduce an electron mediator (methyl viologen,  $\text{MV}^{2+}$ ) with  $\text{SO}_3^{2-}$  as electron donor. Another **catalyst** then uses  $\text{MV}^{+\bullet}$  to produce  $\text{H}_2$ .



methyl viologen,  $\text{MV}^{2+}$



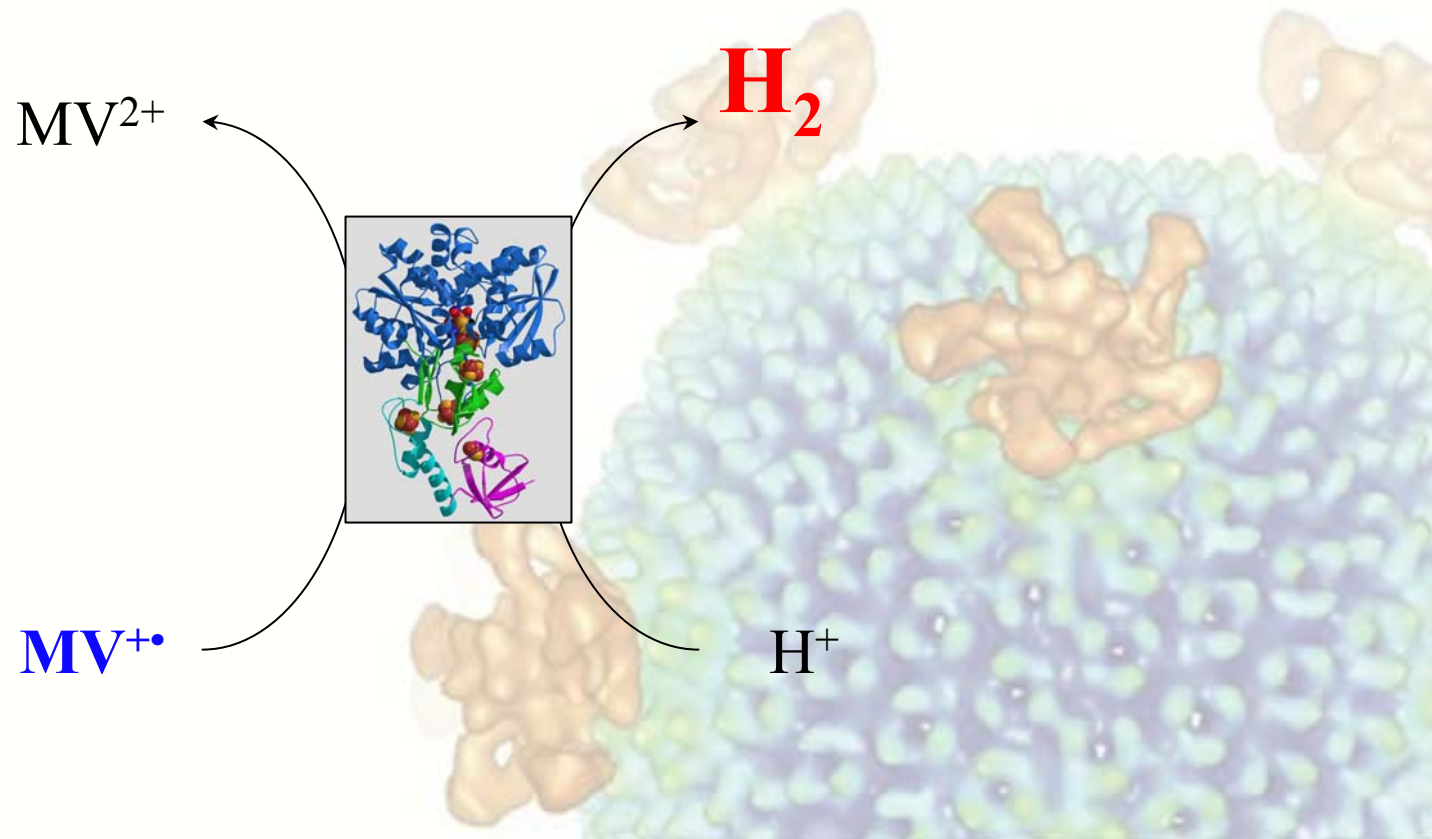
# Enzymatic H<sub>2</sub> Formation



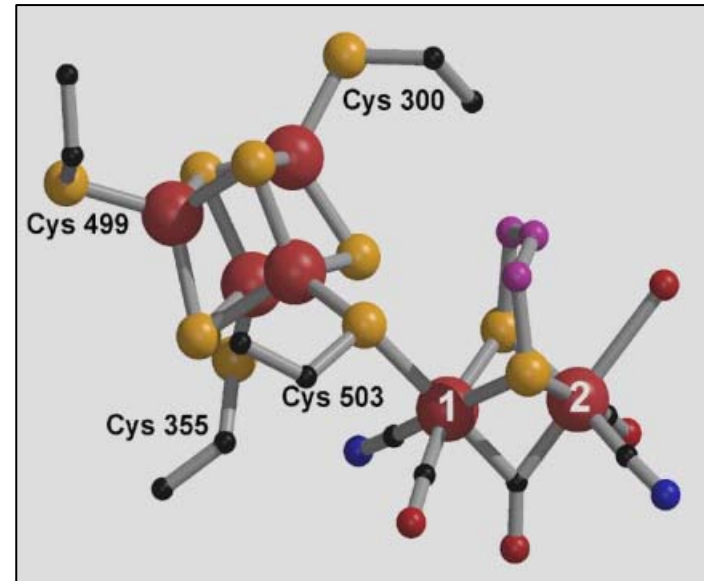
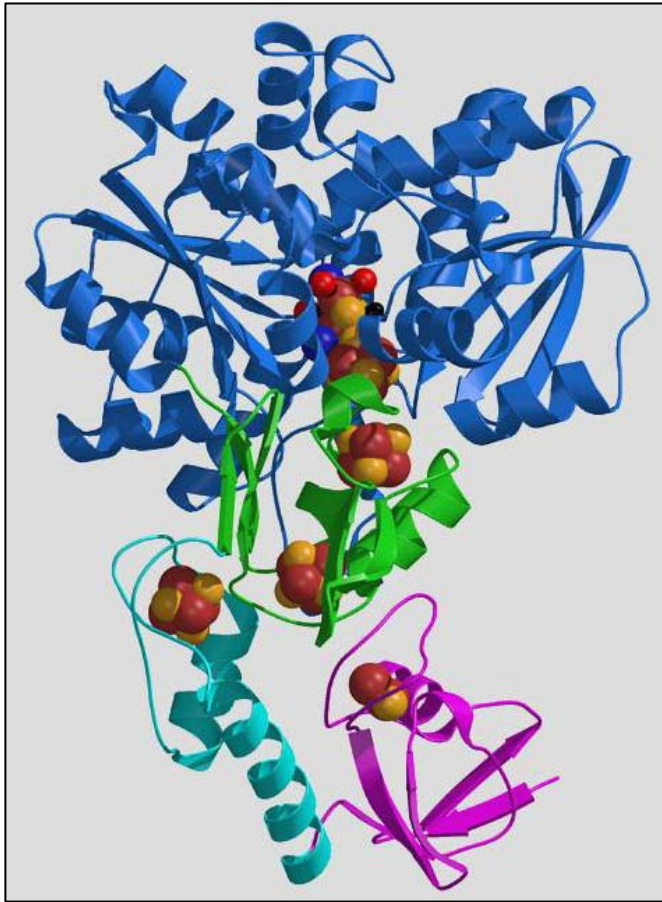
Hydrogenase enzymes

Highly active catalysts (9,000 H<sub>2</sub>/enz/sec)

Utilize MV<sup>+</sup> as reducing equivalents



# ***Biological Hydrogen Production***



## ***Hydrogenase***

Peters et al, *Science* (1998)





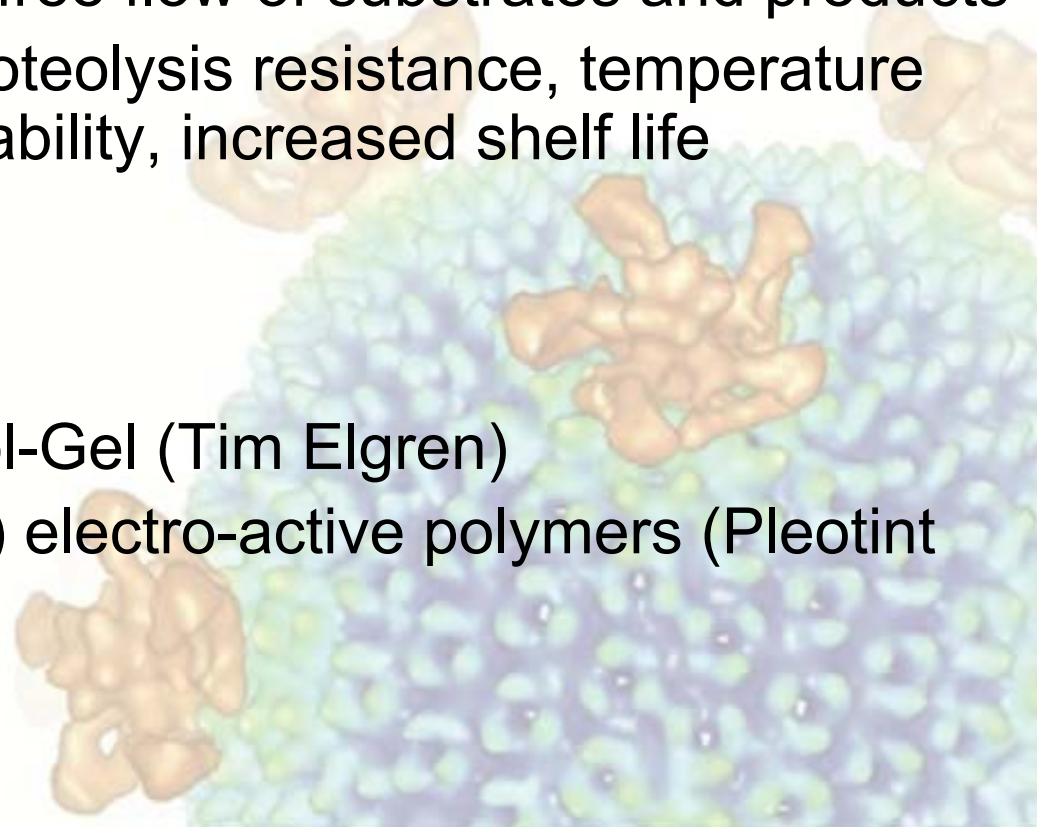
# Hydrogenase Immobilization

## Advantages

- Solid Phase – free flow of substrates and products
- Durability – Proteolysis resistance, temperature stability, pH stability, increased shelf life

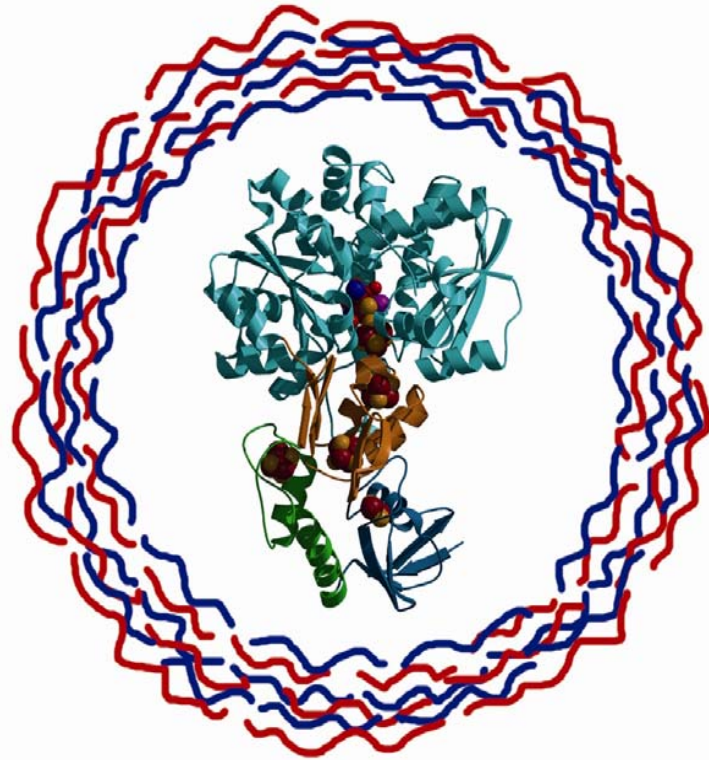
## Approaches

- Silica oxide Sol-Gel (Tim Elgren)
- Poly(viologen) electro-active polymers (Pleotint LLC)

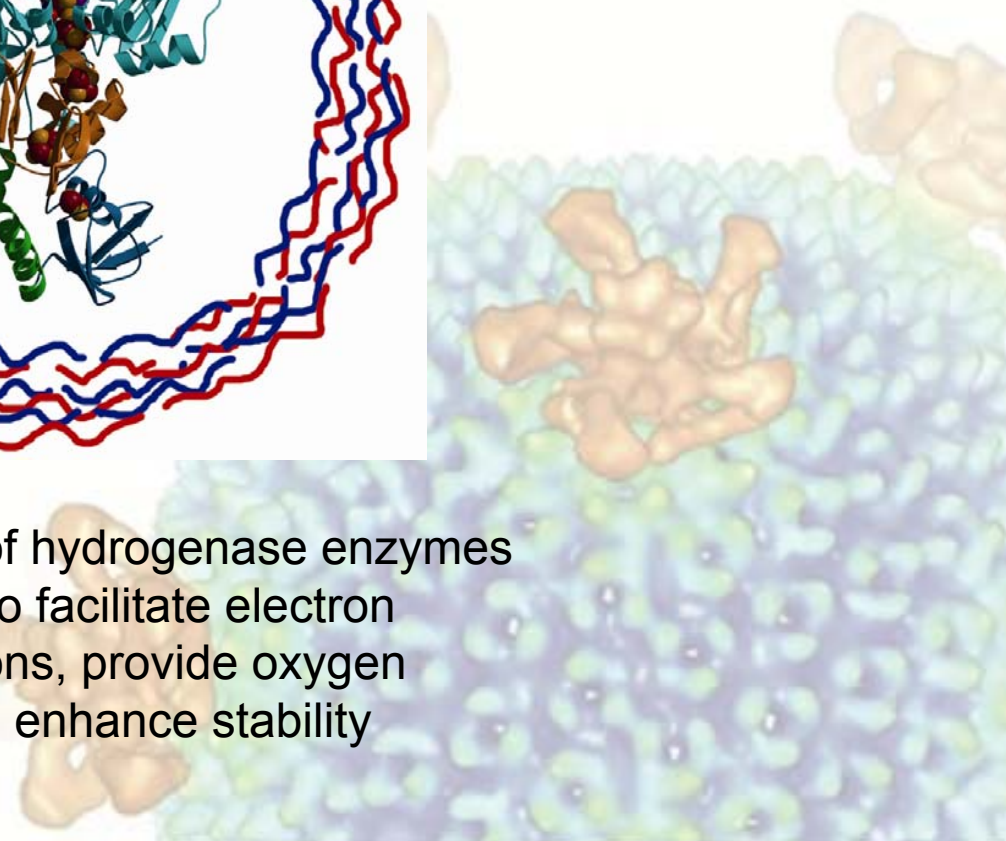




# Encapsulation/Immobilization



Incorporation of hydrogenase enzymes into materials to facilitate electron transfer reactions, provide oxygen protection, and enhance stability



# Procedure for making Sol-Gel hydrogenase materials



Prepared Sol-Gel mixture

*1.57 ml Tetramethyl-ortho-silicate (TMOS)*  
*350  $\mu\text{L}$   $\text{H}_2\text{O}$*   
*11  $\mu\text{L}$  0.04 M HCl*

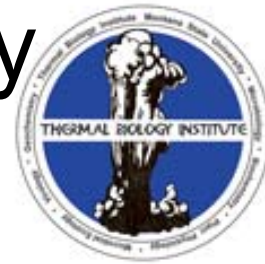
Sonicated solution for 30 min in cold bath  
(with degassing)

Making Sol-Gel hydrogenase materials

*100  $\mu\text{L}$  hydrogenase (100  $\mu\text{g}$  of protein  
in 50 mM Tris- HCl pH 8,0) :*  
*100  $\mu\text{L}$  Sol-Gel mixture*

Polymerization of Sol- Gel material for 3-5 min

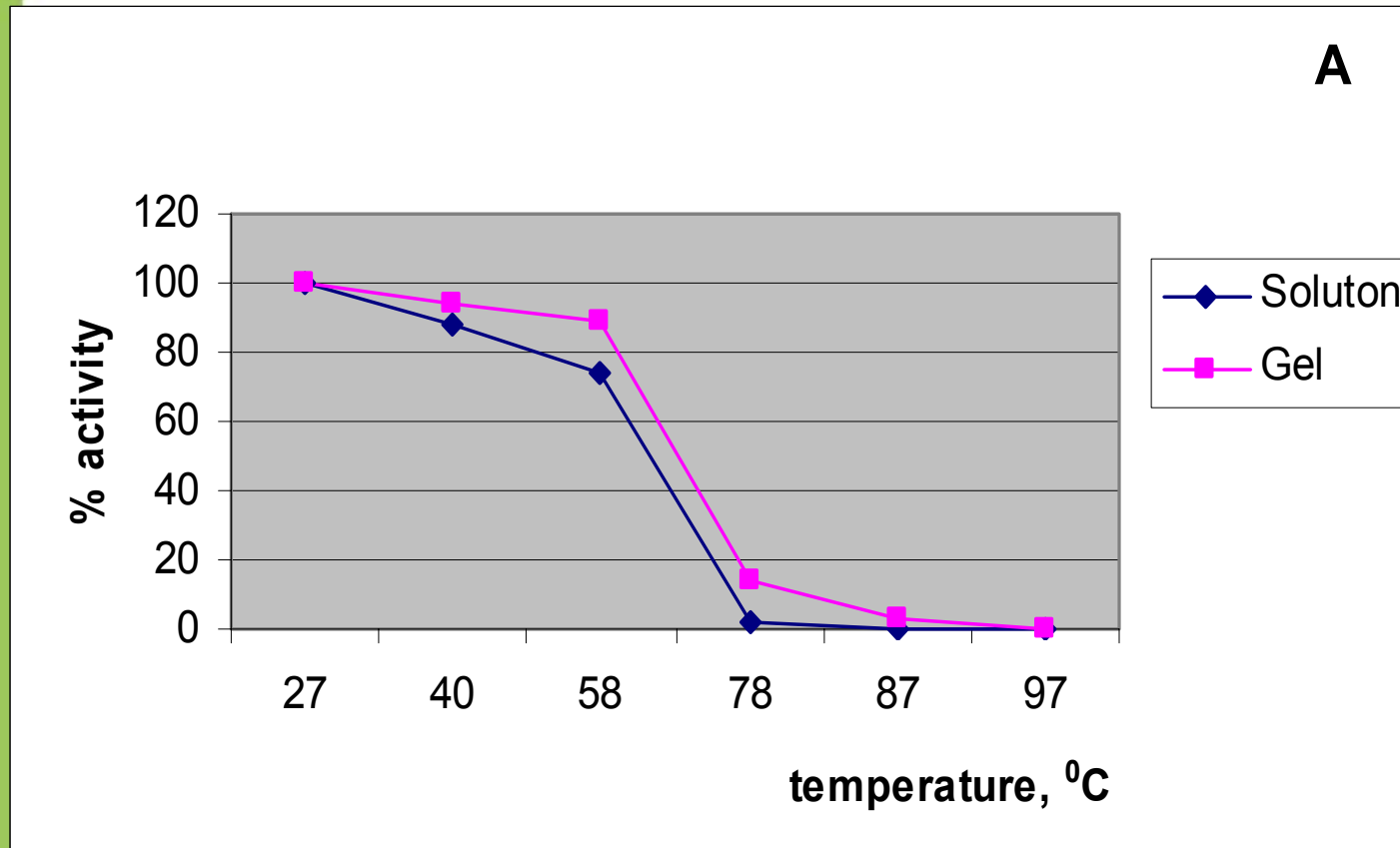
# Recovery of hydrogenase activity encapsulated in Sol-Gel



Hydrogenase		% activity	
		Solution	Sol-Gel
1.	<i>Clostridium pasterianum</i>	100	63.8±15.8
2.	<i>Lamprobacter modestogalophilus</i>	100	67.5±8.8
3.	<i>Thiocapsa roseopersicina</i>	100	70.1±2.5



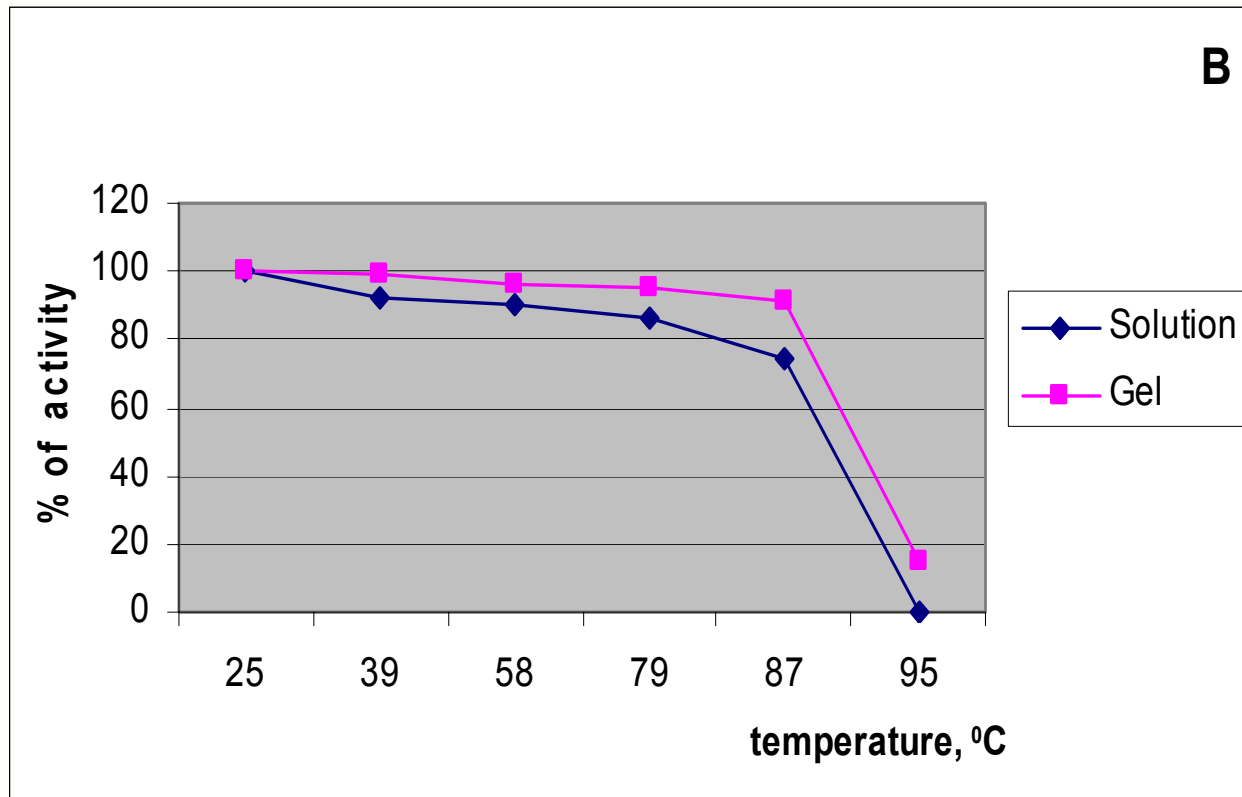
# Thermal stability of hydrogenases encapsulated in Sol-Gel materials



*A - Clostridium pasterianum hydrogenase*



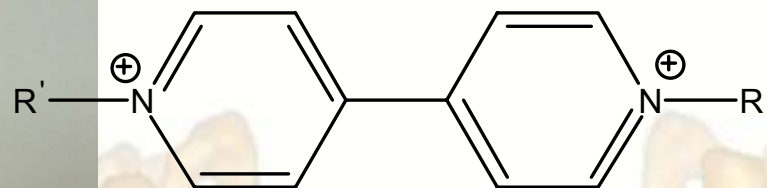
# Thermal stability of hydrogenases encapsulated in Sol-Gel materials



*B - Lamprobacter modestogalophilus hydrogenase*

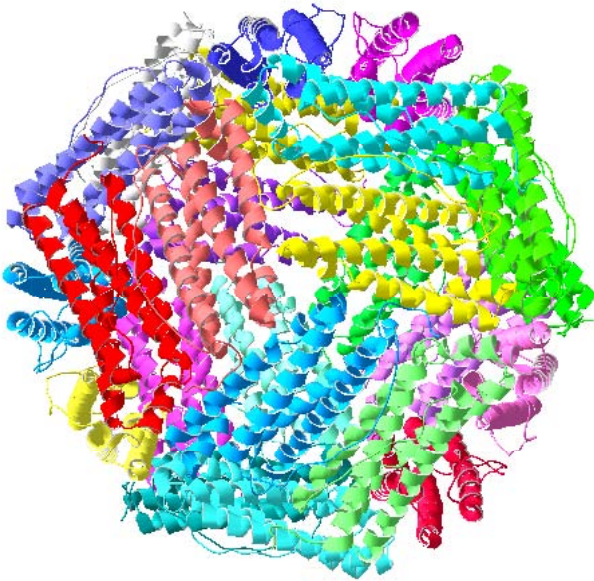


# Association of Redox Mediator with Gel Matrix

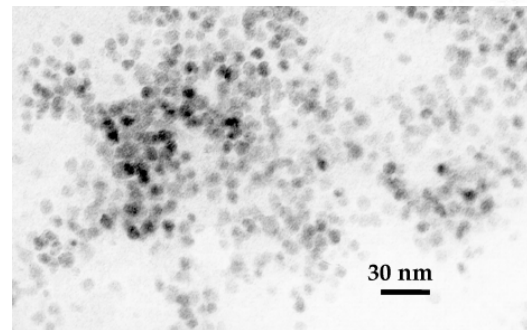
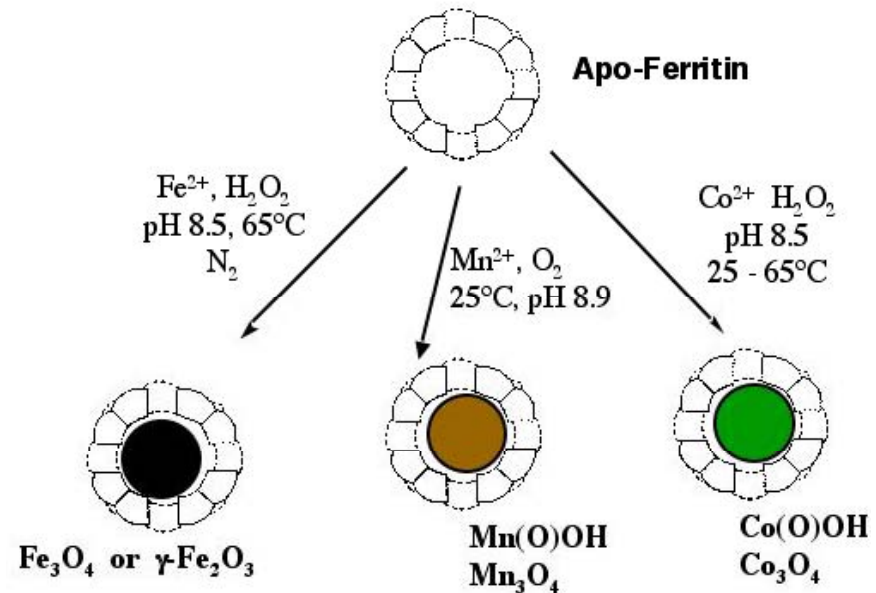


- $\text{R}, \text{R}' = \text{CH}_3$   
 $\text{CH}_2\text{CH}_2\text{CH}_2\text{-SO}_3^-$   
 $\text{CH}_2\text{CH}_2\text{CO}_2^-$   
 $\text{CH}_2\text{CH}_2\text{NH}_3^+$   
 $(\text{CH}_2)_n\text{-PO}_3\text{H}_2$

# Nanoparticle synthesis within the Ferritin Protein Cage



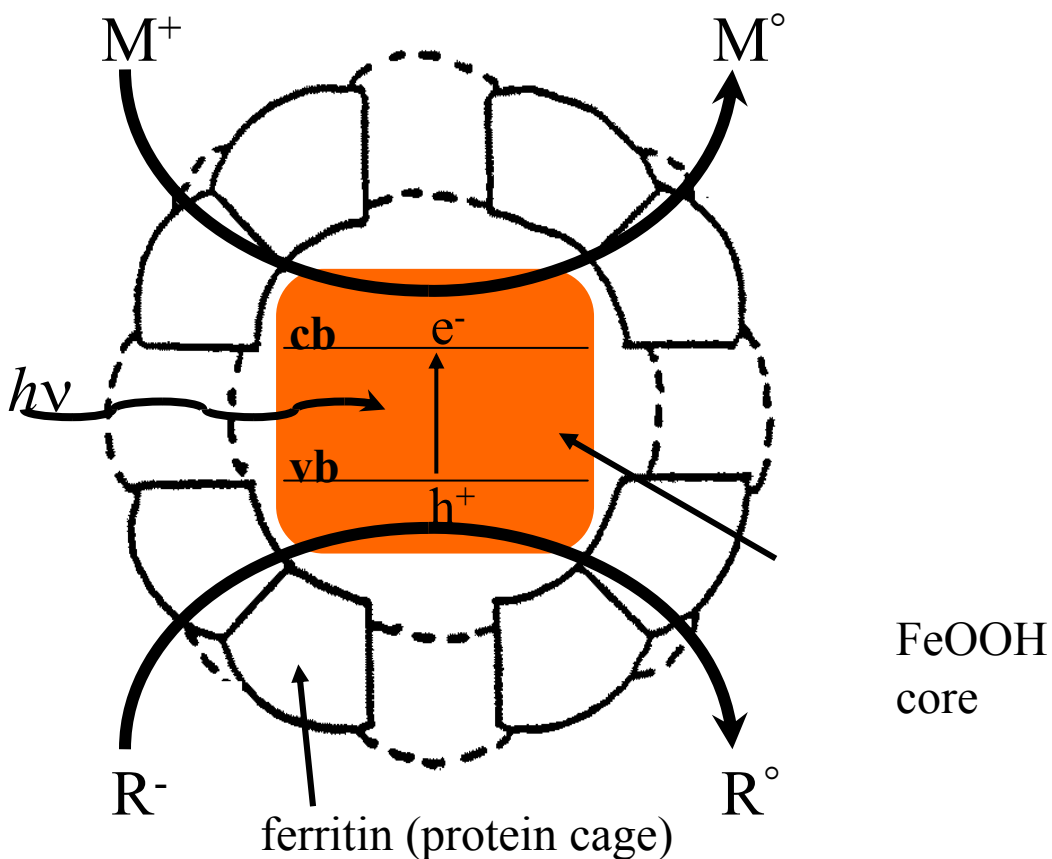
Ferritin protein cage  
24 subunits - 12 nm diam



TEM of metal oxide nanoparticles



# Protein Cage Photocatalysts



Light absorption by ferritin core (FeOOH) causes charge separation oxidizes  $R^-$  and reduces  $M^+$  catalytically.

*Examples:*

- Reduction of  $CrO_4^{2-}$  to Cr(III) using tartrate as electron donor (Kim et al., *Chem. Mater.*, 2002).
- Reduction of Cu(II) to Cu(0) particles using citrate as electron donor (Ensign et al., *Inor. Chem.*, 2004).

**Current: use this photocatalytic system (or an analogue) to reduce  $MV^{2+}$  to  $MV^{+}$  using sulfite as electron donor.**

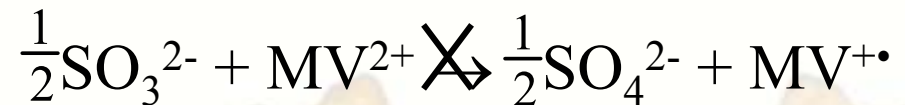


# Thermodynamics and Kinetics

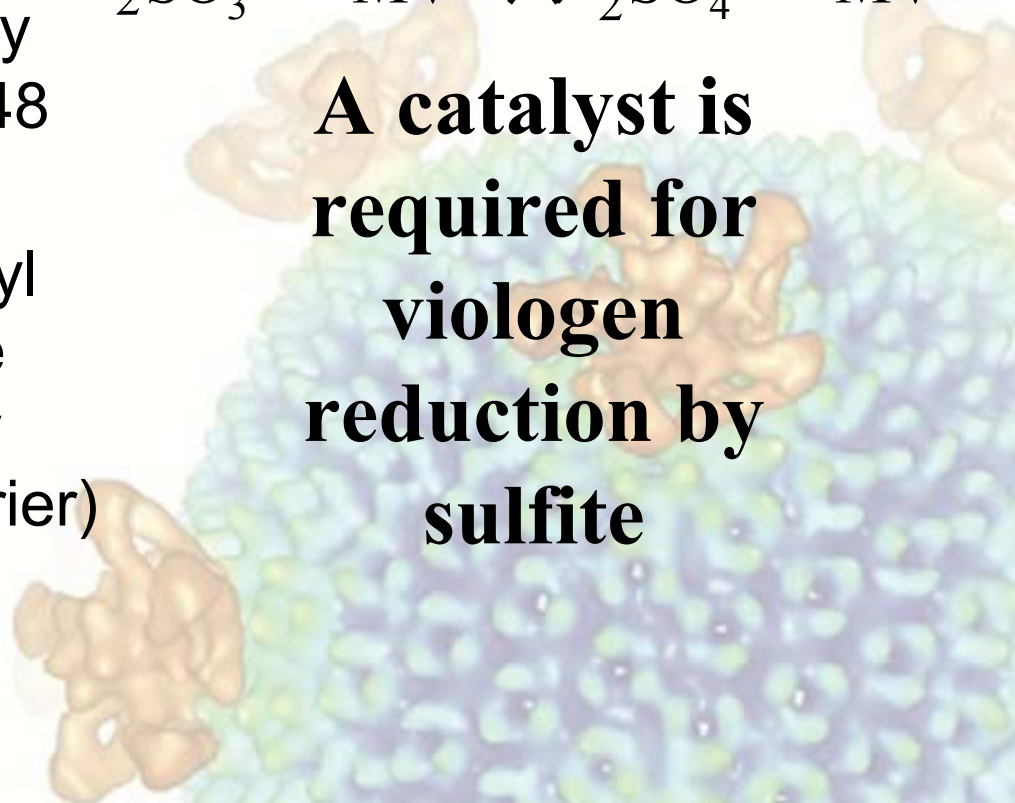


- electron transfer from sulfite to methyl viologen is thermodynamically favorable,  $\Delta G = -48$  kJ/mol
- reduction of methyl viologen by sulfite does not normally occur (kinetic barrier)

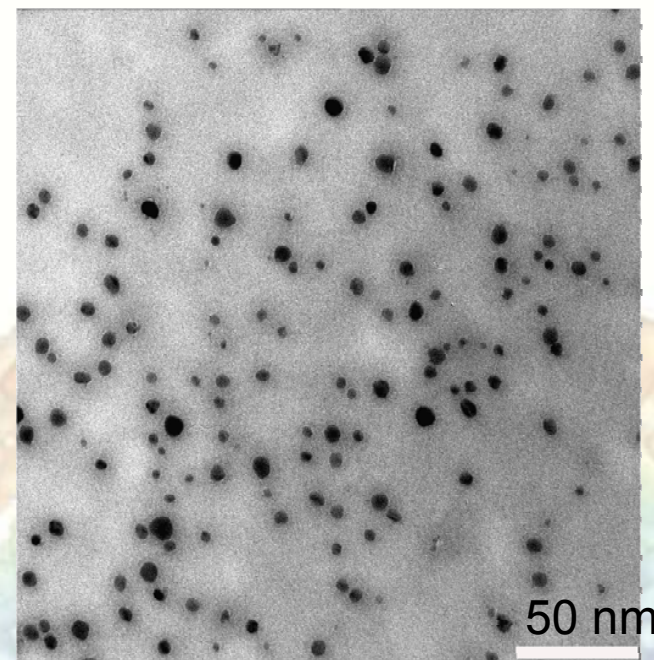
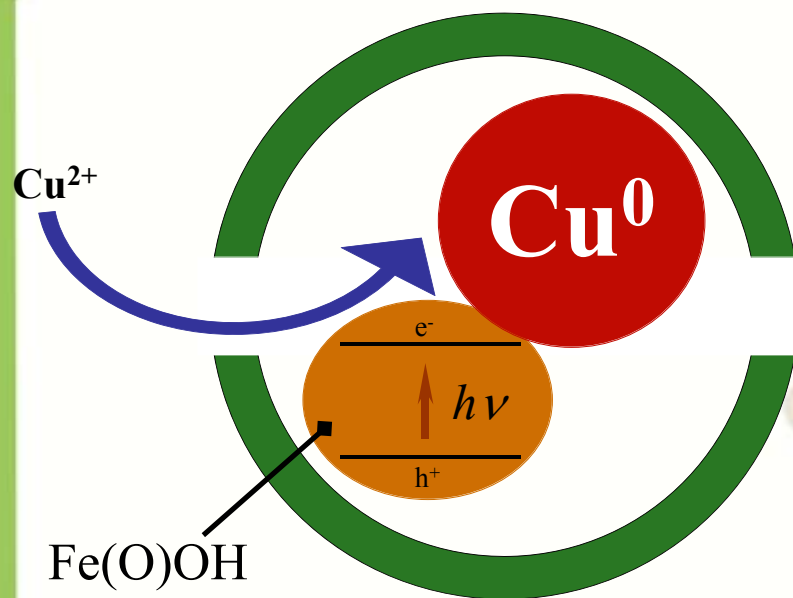
viologen reduction



**A catalyst is required for viologen reduction by sulfite**



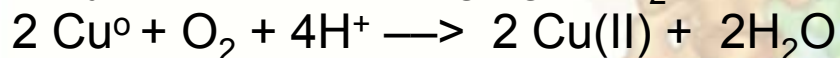
# Photoreduction of Cu(II)



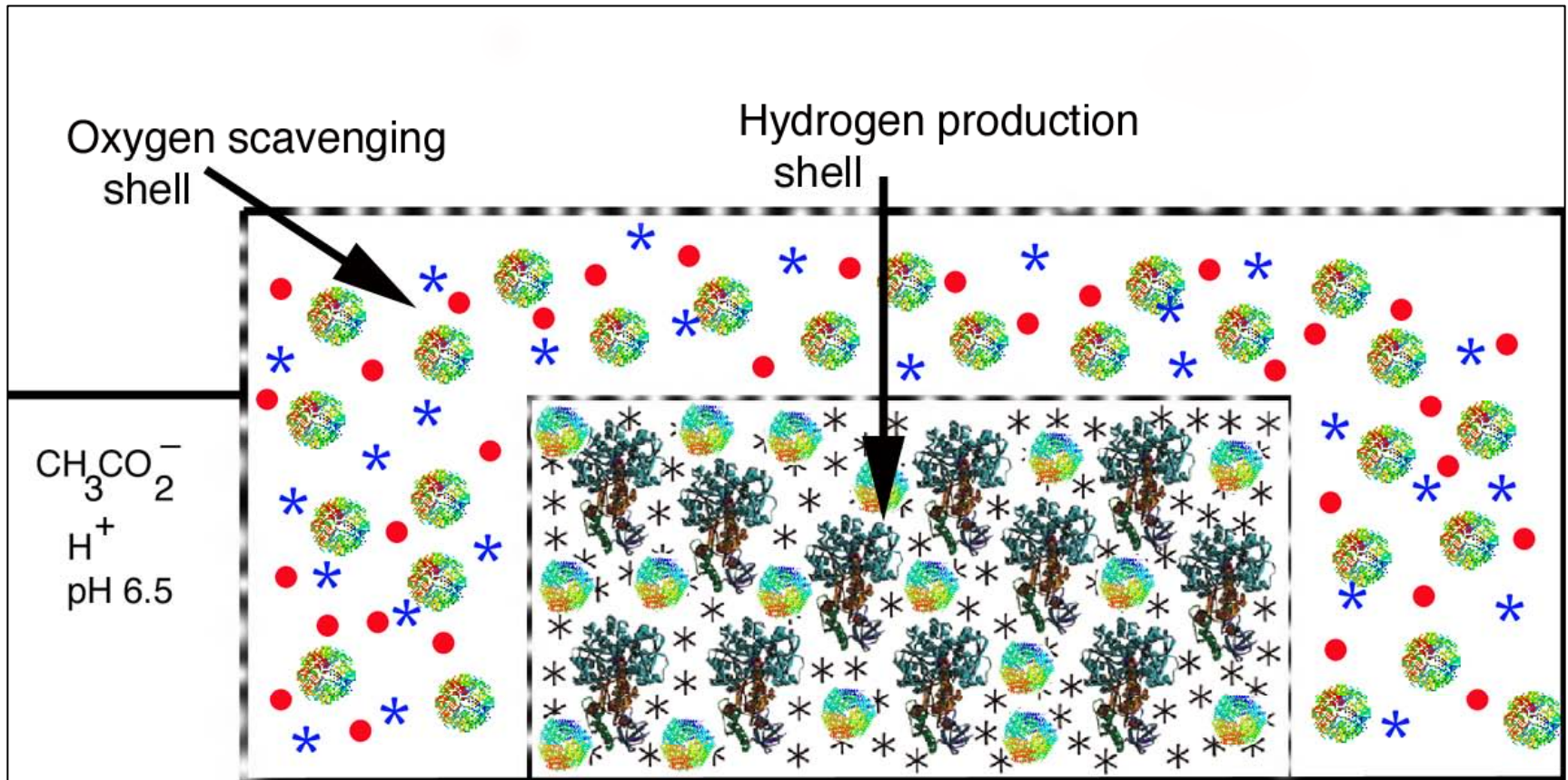
TEM of Ferritin encapsulated  
Cu nanoparticles

Photoreduction of Cu(II) to form protein encapsulated  $\text{Cu}^0$  nanoparticles

Very efficient scavenging of  $\text{O}_2$  from the media



# Long-Term Goal – Device for photocatalytic hydrogen production – composite materials (nanoparticles and hydrogenase enzymes)





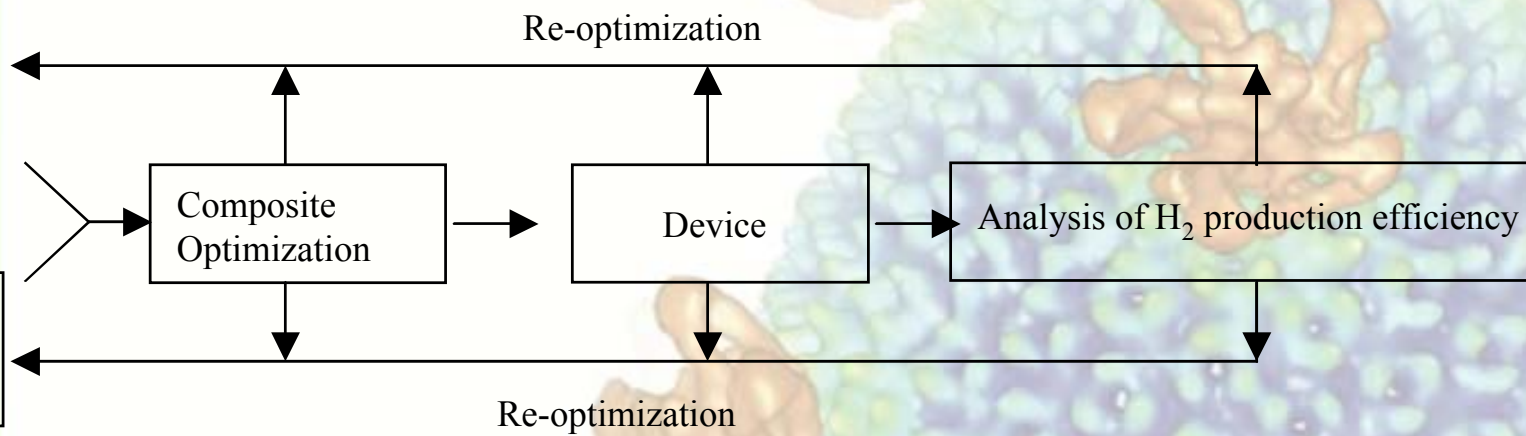
# Future Goals



Development of device prototype  
demonstration of light harvesting

Hydrogenase  
Optimization

Semiconductor  
Nanoparticle  
Optimization



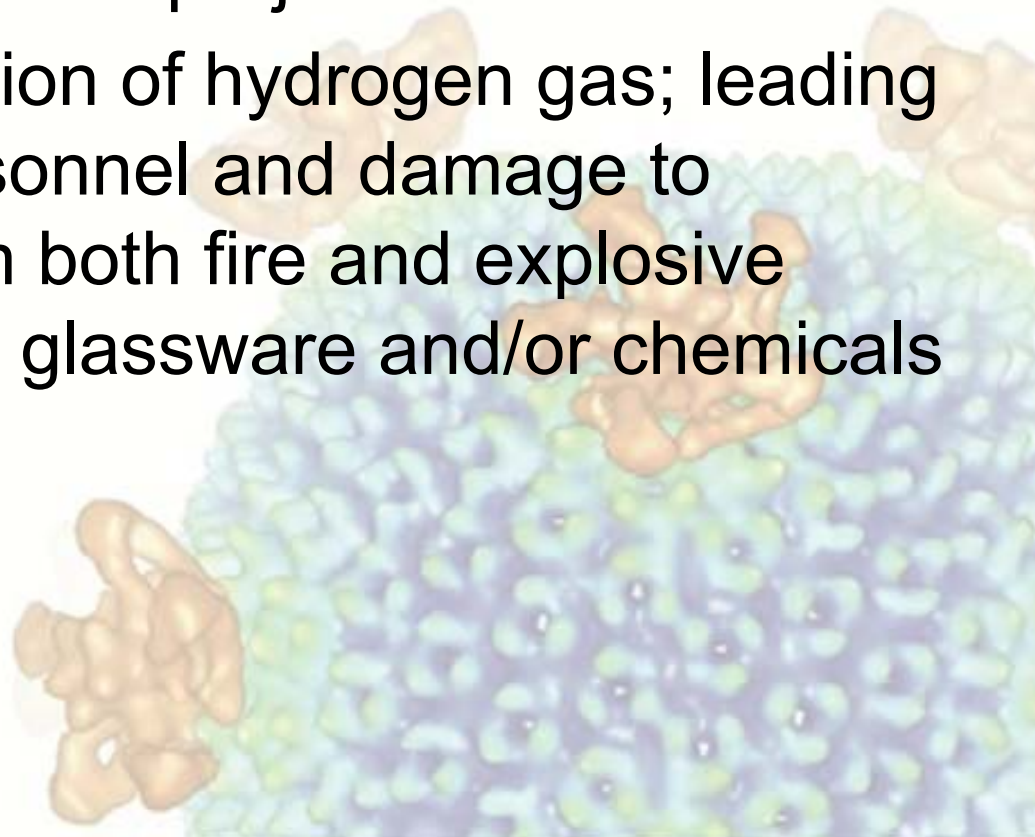


# Hydrogen Safety



The most significant hydrogen hazard associated with this project is:

Accidental ignition of hydrogen gas; leading to injury of personnel and damage to equipment from both fire and explosive debris such as: glassware and/or chemicals





# Hydrogen Safety

Our approach to deal with this hazard is:

Follow lab protocol of wearing safety glasses, gloves

Keep glove box  $H_2$  level below 3%

Vent gases in fume hood

Keep away from open flame and flammable chemicals

Keep quantity of  $H_2$  production to a minimum

In event of accidental explosion contact

Jeff Shada, Safety and Risk Management,  
Advanced Tech Park, 406-994-2711