

# Hydrogen from Water in a Novel Recombinant Oxygen-Tolerant Cyanobacterial System

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Energy Laboratory

**Project ID**  
**PD039**

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# Overview

## Timeline

- Project start date: 5-01-2005
- Project end date: 1-30-2014
- Percent complete: 91%

## Budget

- Total project funding
  - DOE share: \$2.019M for JCVI
  - DOE share: \$1.26M for NREL
  - JCVI cost-share: \$820K
- Funding received for FY12
  - \$200K for JCVI
- Planned funding for FY13
  - \$150K for JCVI

## Barriers

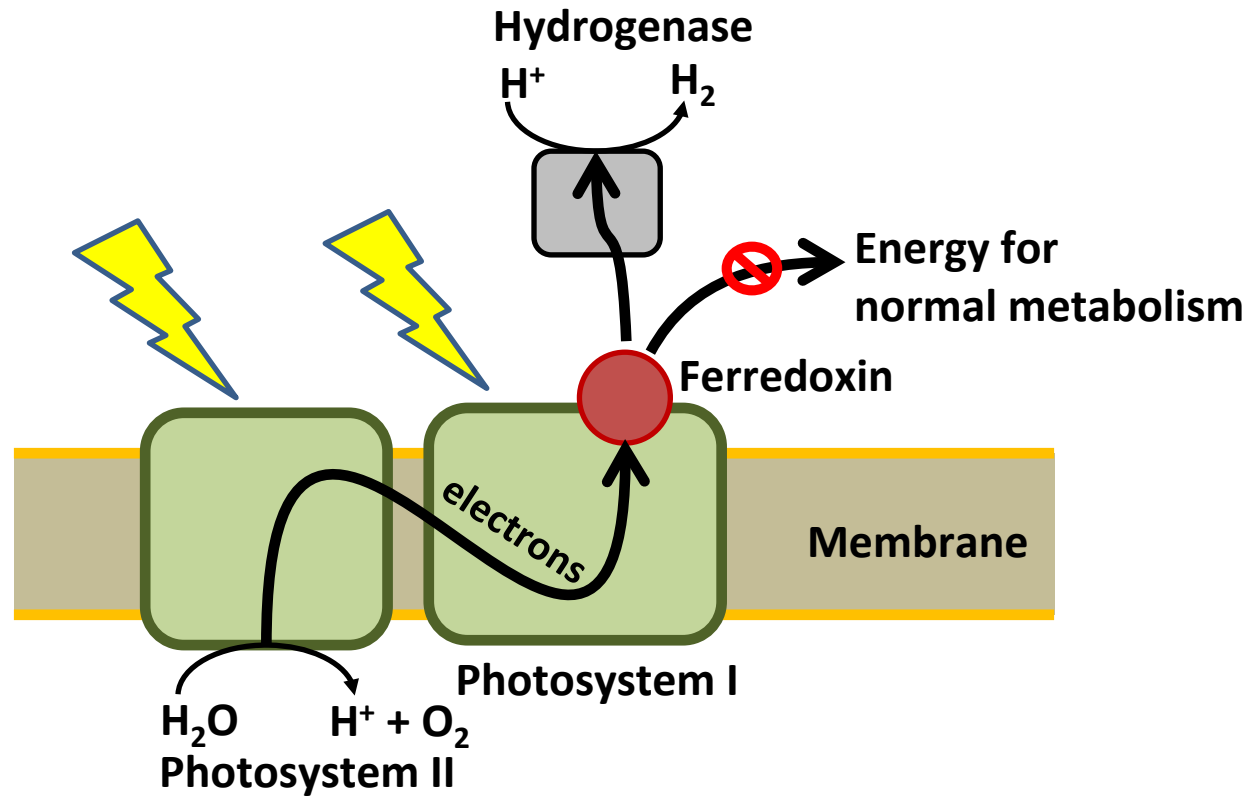
- Barriers addressed
  - Biological Hydrogen Production Barrier AP: Oxygen Accumulation

## Partners

- National Renewable Energy Laboratory

# Relevance:

**Develop an O<sub>2</sub>-tolerant cyanobacterial system for continuous light-driven H<sub>2</sub> production from water**



Barrier AP: O <sub>2</sub> Accumulation	2011 Status	2015 Target	2020 Target
Duration of continuous photoproduction in full sunlight	2 min	30 min	4 h

# Approach: Milestones and Go/No Go

## Task 1. Engineering known hydrogenases

Month/Year	Milestone	% Comp
Sept-11	Purify hydrogenase and verify functionality in O <sub>2</sub>	100%
Dec-10	Determine electron mediator requirement	100%
Sept-11	Verify hydrogenase activity in cyanobacteria <i>in vivo</i> and ability to make H <sub>2</sub> from water	100%

## Task 2. Discovery and engineering of new hydrogenases

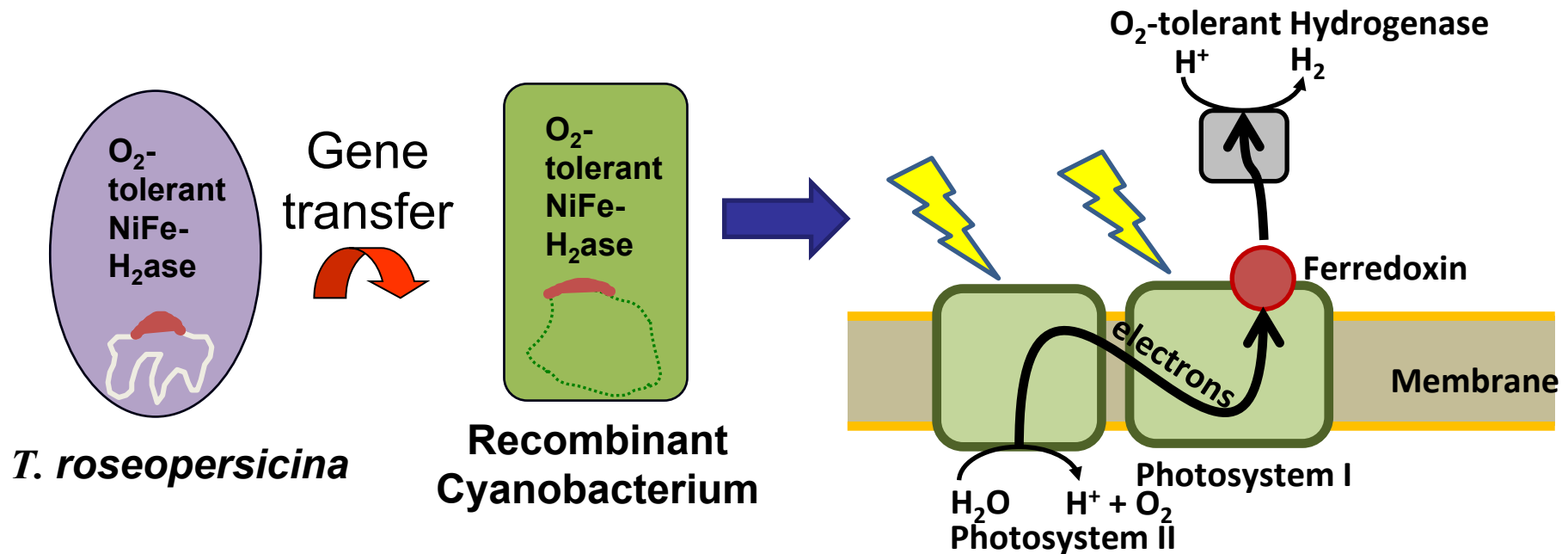
Month/Year	Milestone	% Comp
Sept-10	Identify novel hydrogenases from environment and transfer to cyanobacteria	100%
Aug-11	Construct a cyanobacterial hybrid to express an active environmental hydrogenase	100%
Apr-13	Increase activity of HynSL hydrogenase in cyanobacteria to give 100-fold increase in specific activity.	30%
Nov-13	Improve hydrogenase-ferredoxin (Fd) electron transfer to enable 25-fold better Fd docking to the hydrogenase.	20%
Jan-14	Measure light-dependent H <sub>2</sub> production in modified strains.	5%

**Go/No Go Decision: Due Jan-13, Achieved Nov-12**

Demonstrate 5x increase hydrogenase activity from environmental H<sub>2</sub>ase in cyanobacteria as measured by *in vitro* H<sub>2</sub> evolution assay.

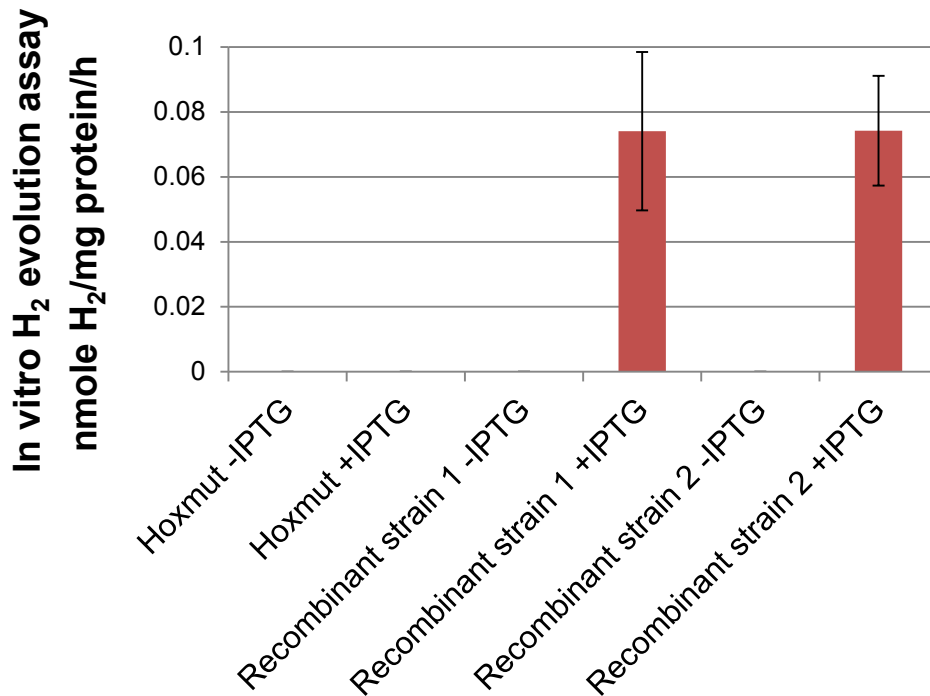
# Task 1: Technical Approach

Transferring a known O<sub>2</sub>-tolerant NiFe-hydrogenase from *T. roseopersicina* into cyanobacterium *Synechococcus* sp. PCC 7942



# Task 1: Technical Approach

## Review from previous AMR:

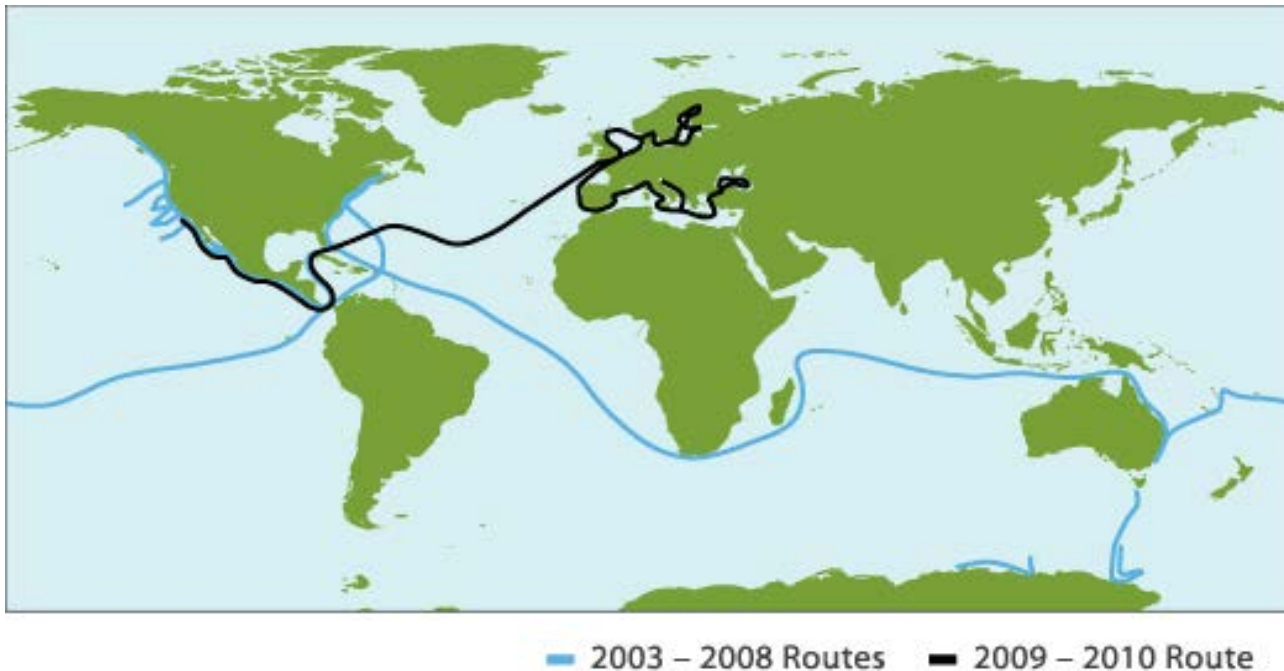


- **Completed milestone “Construct cyanobacterial hybrid to express active *Thiocapsa* hydrogenase” (8/11).**
- Effort in FY2011-13 focused on increasing expression of heterologous hydrogenase using environmentally-derived hydrogenase as model.

- JCVI approach is complementary to that of NREL in harnessing Nature’s O<sub>2</sub>-tolerant hydrogenases and their transfer into two model cyanobacteria.

# Task 2: Technical Approach

**Task 2.** Identifying novel O<sub>2</sub>-tolerant hydrogenases through metagenomic analysis of marine microbes in the global ocean and transferring the hydrogenases into cyanobacteria



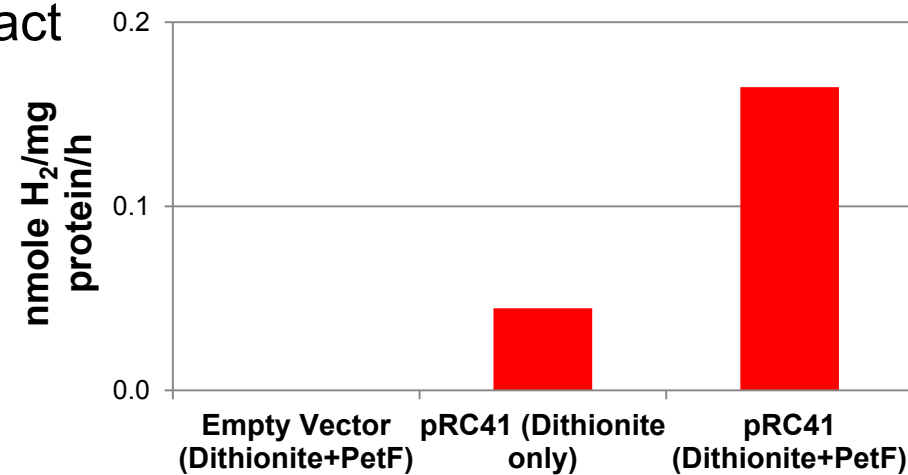
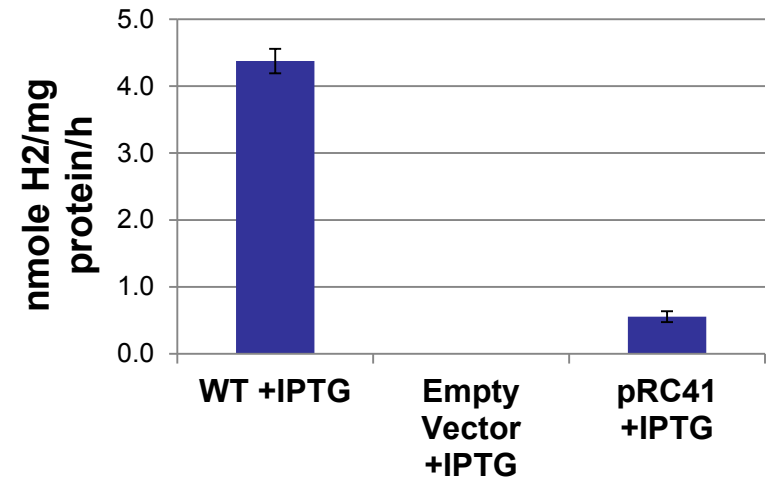
**Sorcerer II Expedition: Global Ocean Sampling Project (Funded as cost share at no expense to DOE)**

- This approach is complementary to two approaches in the Task 1 about harnessing nature's O<sub>2</sub>-tolerant hydrogenases and their transfer into cyanobacteria.

# Task 2: Technical Approach

- Previously, we reported transfer of hydrogenase from environmental DNA to *Synechococcus* PCC 7942 (Top figure, Milestone reached 4/10)
- Environmental hydrogenase is more thermostable and O<sub>2</sub>-tolerant than *Thiocapsa* HynSL.
- Cyanobacterial ferredoxin (PetF\*) can act as an electron mediator to the environmental hydrogenase in *E. coli* crude extract (Bottom figure, Milestone “Determine Electron Mediator Requirement” reached 12/10)

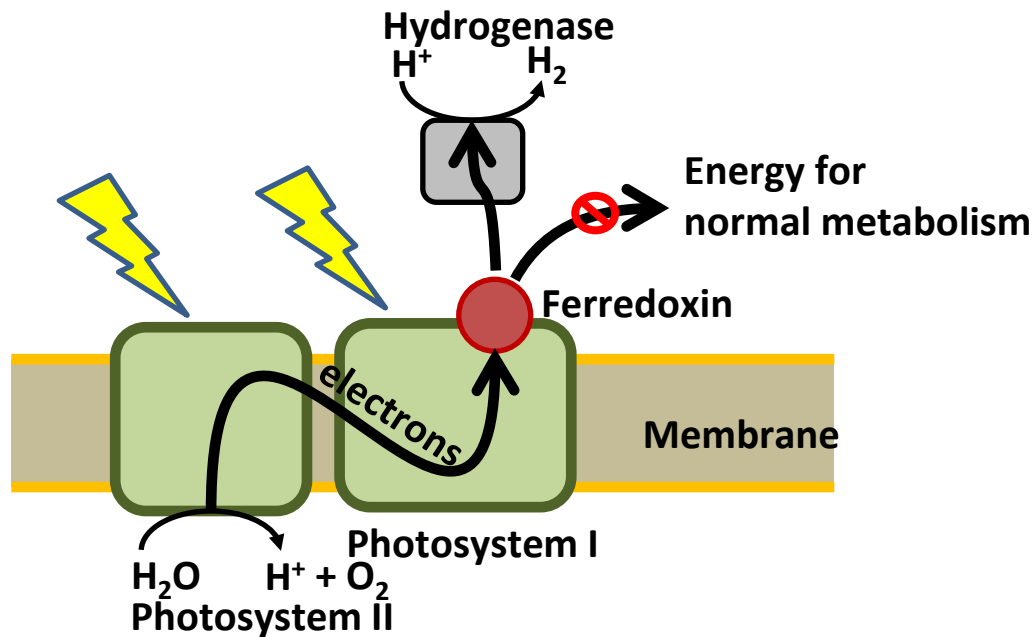
\* PetF cloned and purified by NREL and provided to JCVI through our collaboration





# 2 Current Approaches to Improve System

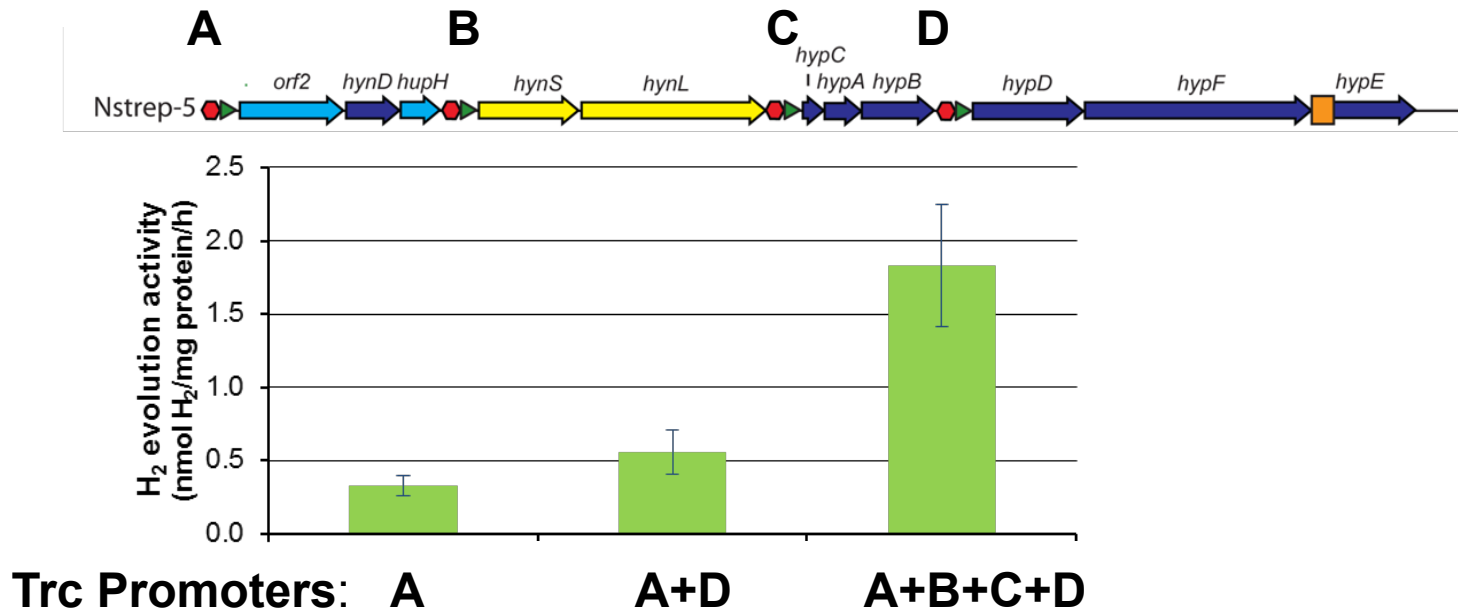
1. Improve expression and activity of hydrogenase in cyanobacteria (Task 2.4)
2. Improve hydrogenase-ferredoxin interaction (Task 2.5)



# Task 2: Technical Approach

## Review from last year's AMR:

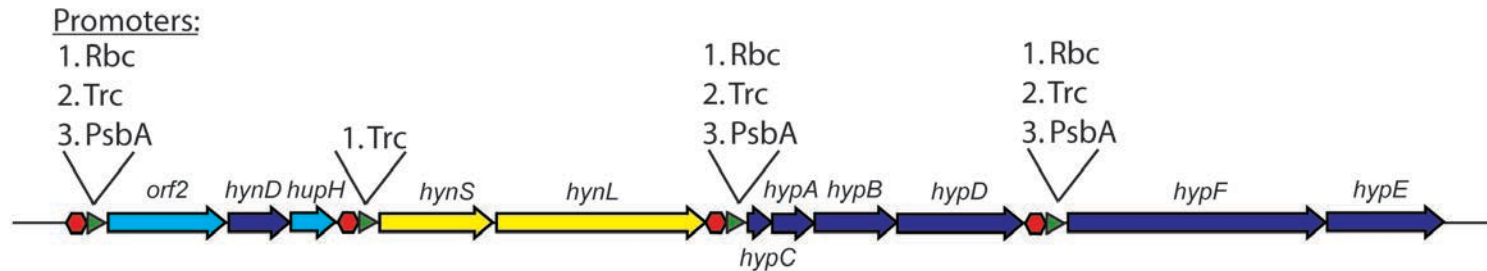
- More Trc promoters increases hydrogenase expression
- Ratios of maturation proteins may be important for maturation efficiency
  - Promoters at A+D led to more “mature” HynL but no increase in activity



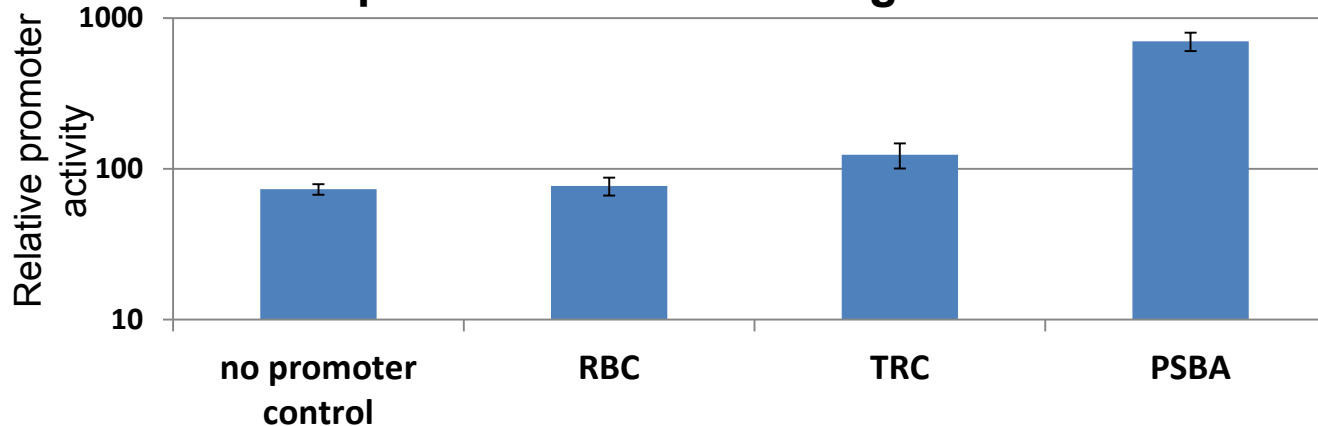
# Task 2: Technical Accomplishments

Testing promoter placement, strength, and frequency

- Plan of promoter placement = 27 different variants



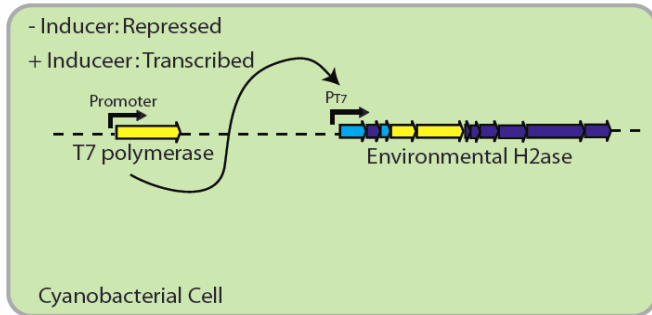
- Relative activities of promoters we are using



**On track to complete Milestone “Increase activity of HynSL hydrogenase in cyanobacteria to give 100-fold increase in specific activity.”**

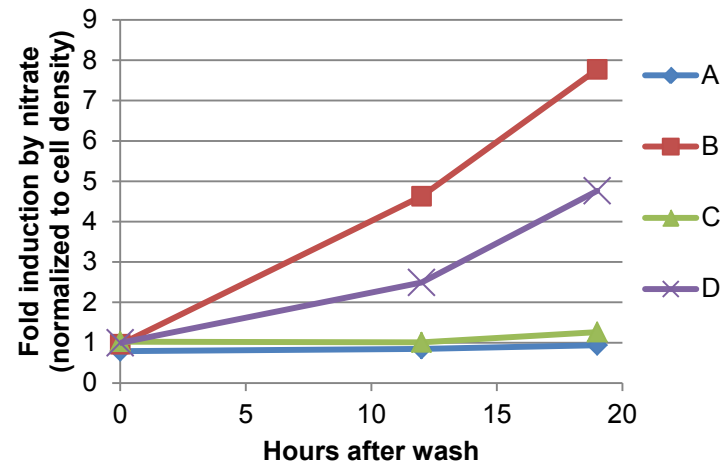
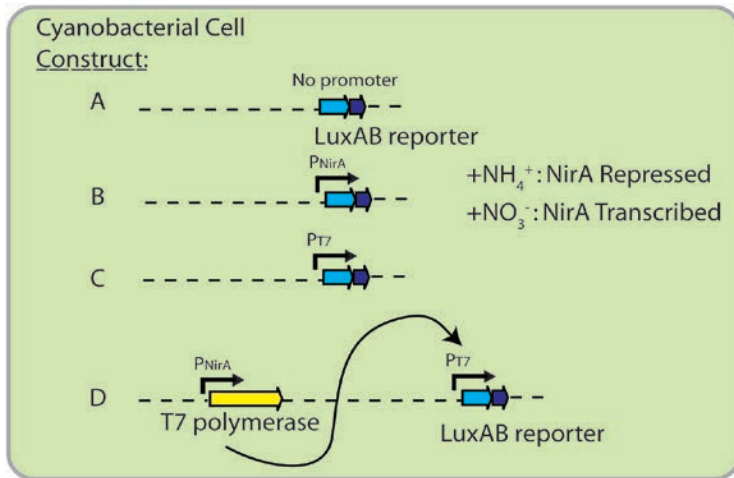
# Task 2: Technical Accomplishments

Engineer T7 polymerase strategy for hydrogenase expression.



## Potential Advantages of T7 polymerase:

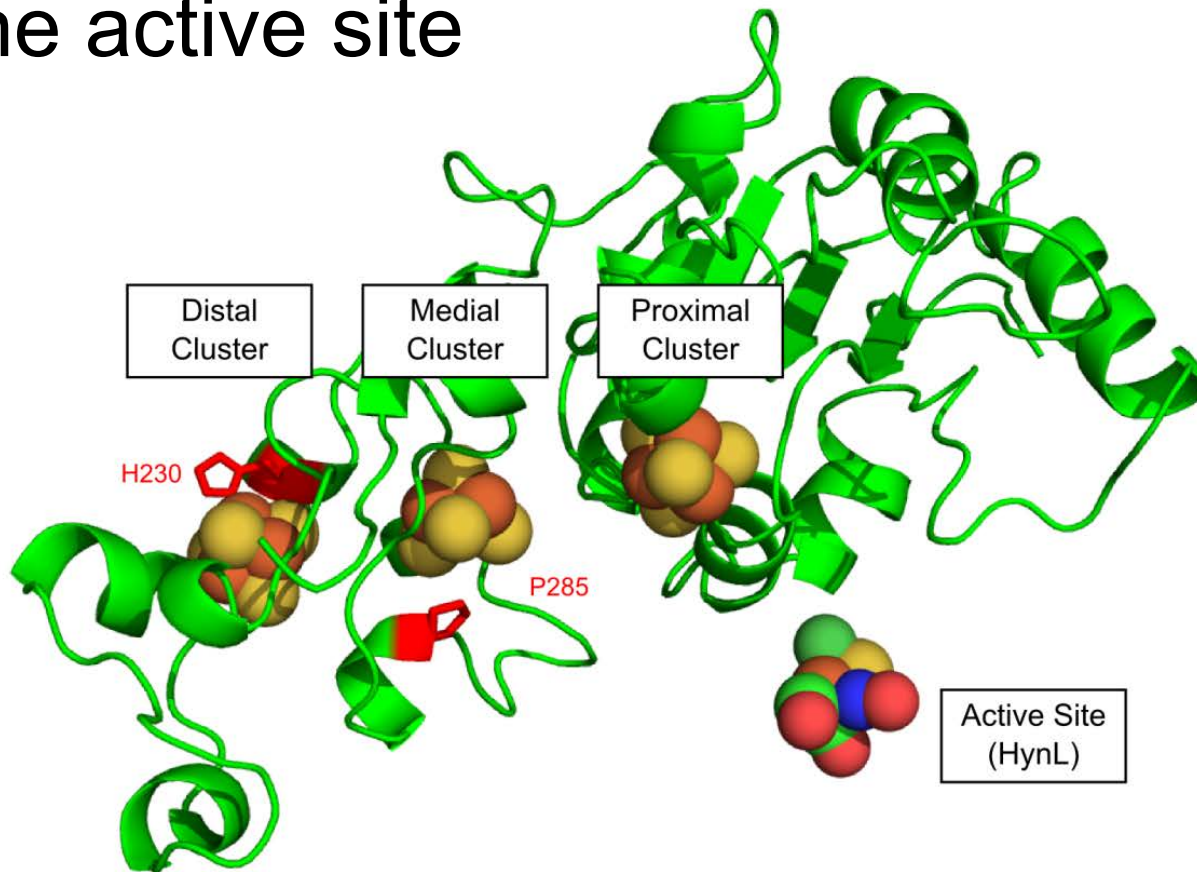
- Tighter control of transcription
- Lower frequency of early termination, may function better with long transcripts.



**Progress toward Milestone “Increase activity of HynSL hydrogenase in cyanobacteria to give 100-fold increase in specific activity.” (4/13).**

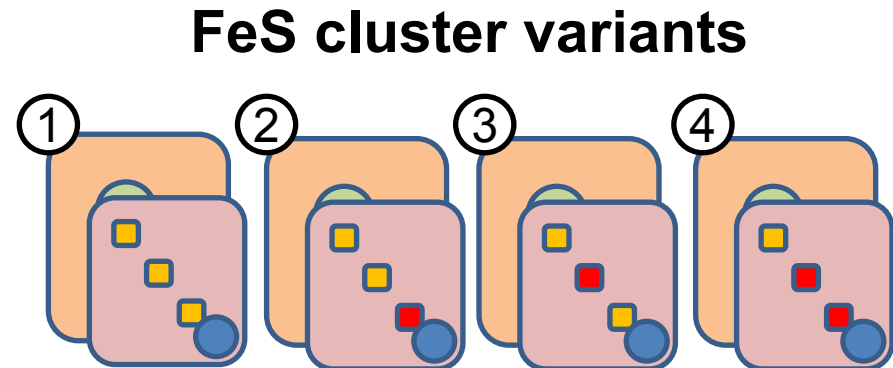
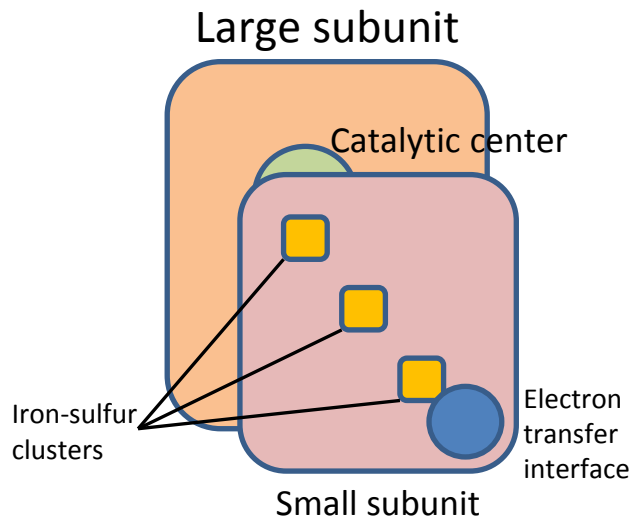
# Task 2: Technical Approach

- FeS clusters form a molecular wire to/from the active site



# Task 2: Technical Approach

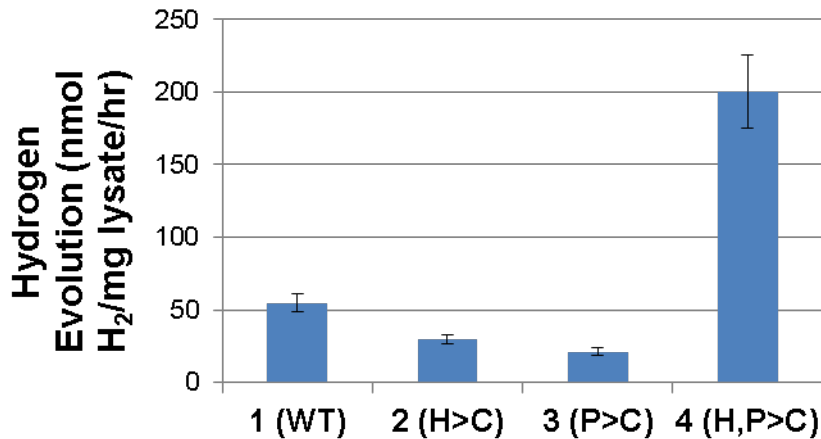
Point mutants alter electrochemistry of the hydrogenase small subunit “molecular wire”



Tests the potential to further modify the environmental hydrogenase to favor H<sub>2</sub> production *in vivo* using PetF as an electron mediator.

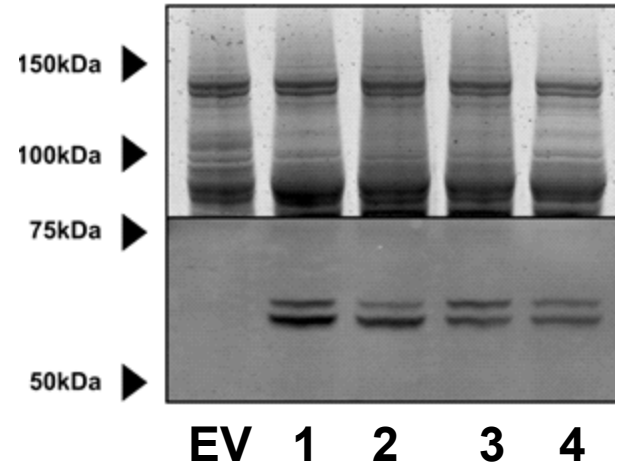
# Task 2: Technical Approach

Review from last year's AMR: Double-substituted HynS yields increased evolution activity relative to uptake



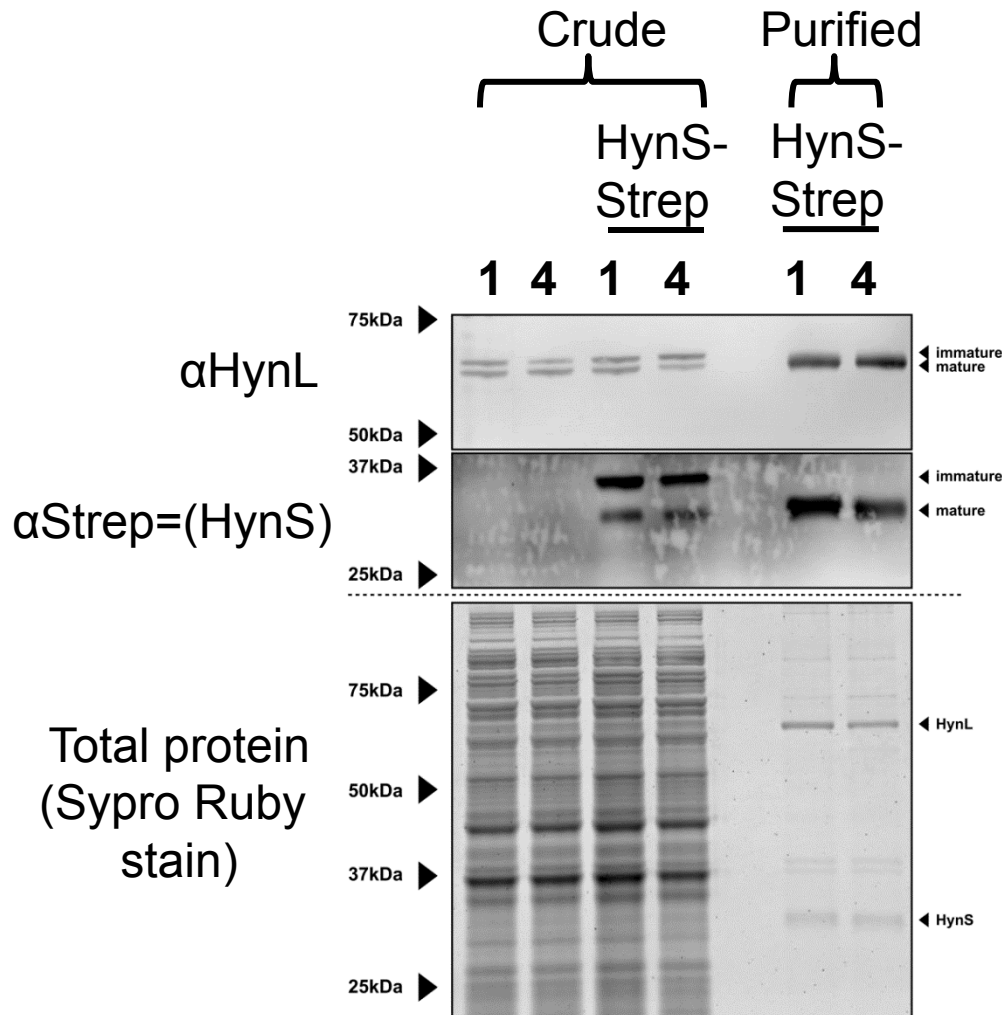
Total protein  
(Sypro<sup>®</sup> Ruby  
stain)

$\alpha$ HynL



# Task 2: Technical Accomplishments

**Increased H<sub>2</sub> evolution in construct 4 is also observed in purified protein**



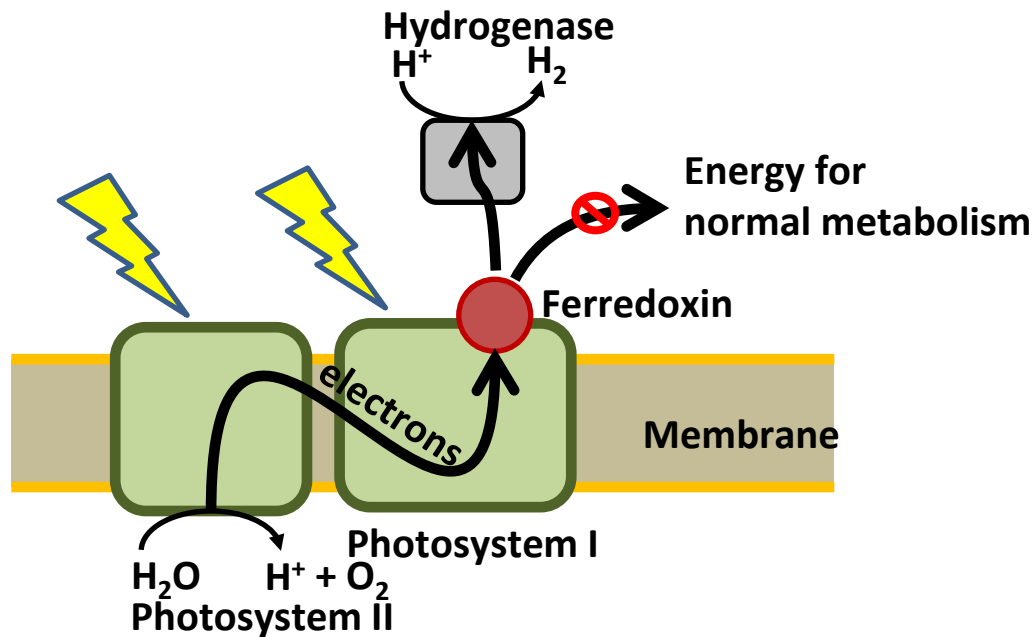
	Specific activity (nmole H <sub>2</sub> produced/mg protein/h)	Fold purification
Construct 1 (crude)	369	
Construct 1 (purified)	41,450	112
Construct 4 (crude)	765	
Construct 4 (purified)	134,214	176

**Completed Milestone 2.5.1 “Develop an affinity-tagged purification system for HynSL” (08/12).**

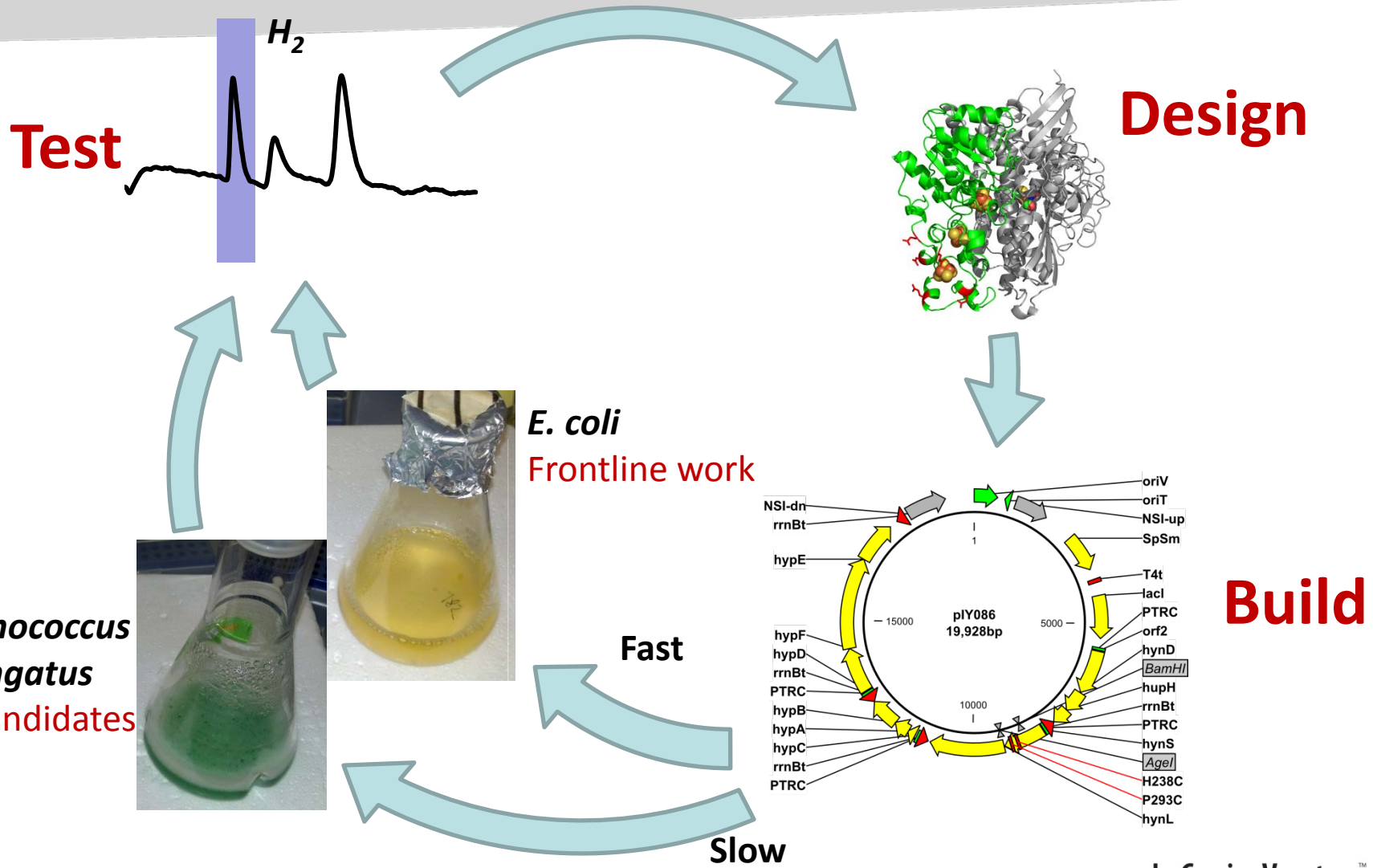


# 2 Current Approaches to Improve System

1. Improve expression and activity of hydrogenase in cyanobacteria (Task 2.4)
2. Improve hydrogenase-ferredoxin interaction (Task 2.5)

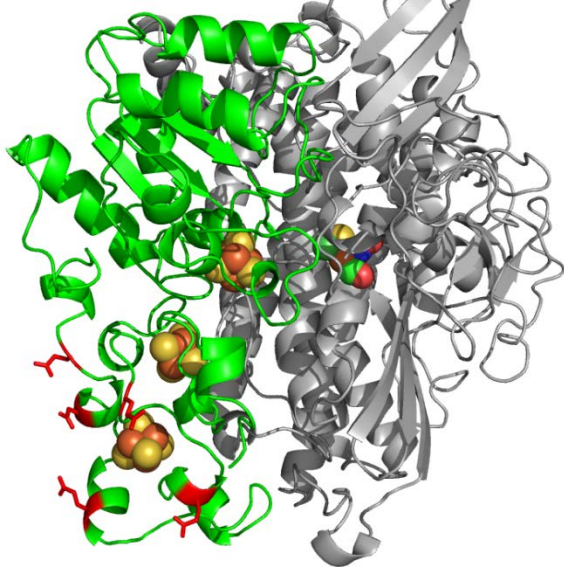


# Engineering cycle for improving H2ase activity

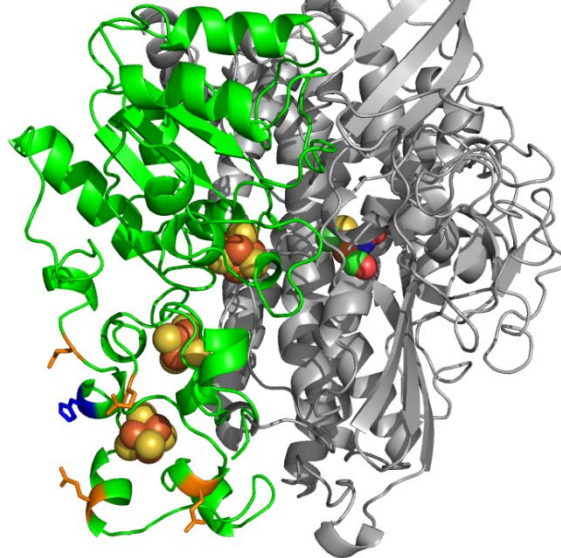


# Task 2: Technical Accomplishments

Wild type



Mutant



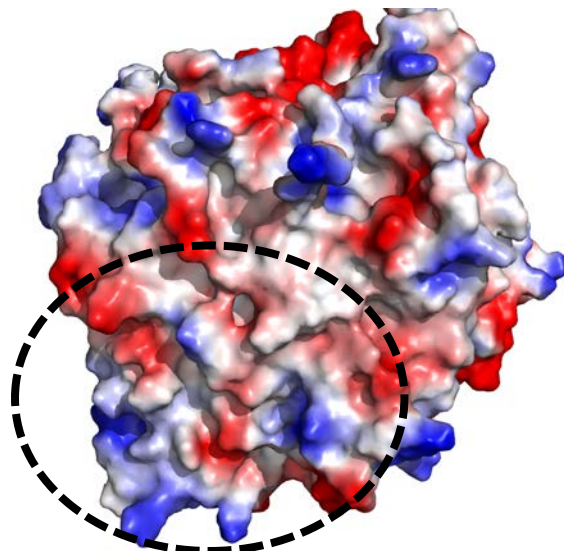
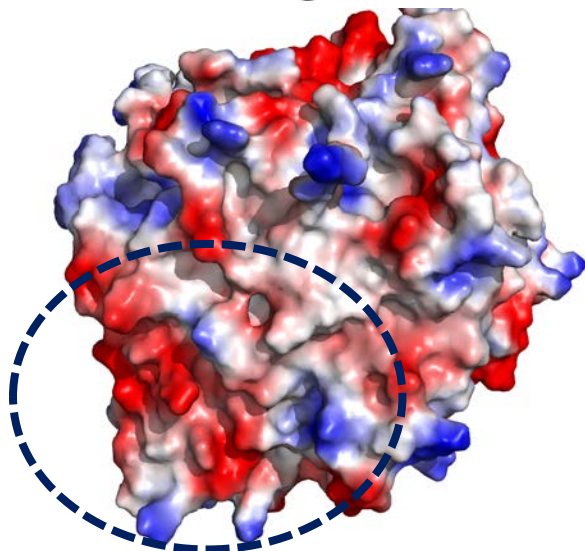
HynS mutants with altered ferredoxin binding sites retain activity

Key:

Red = negative charge

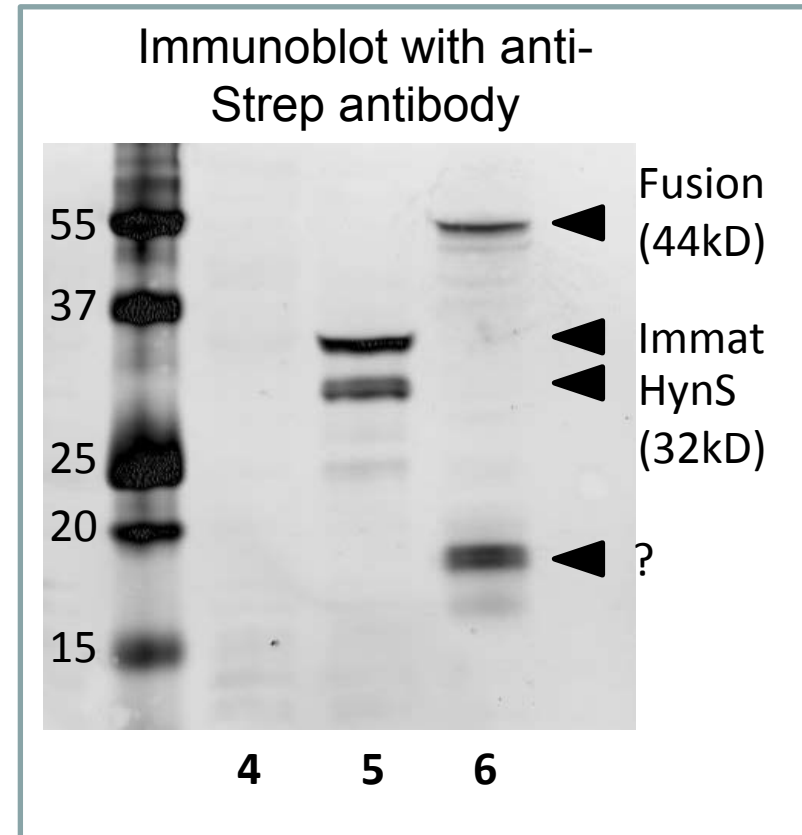
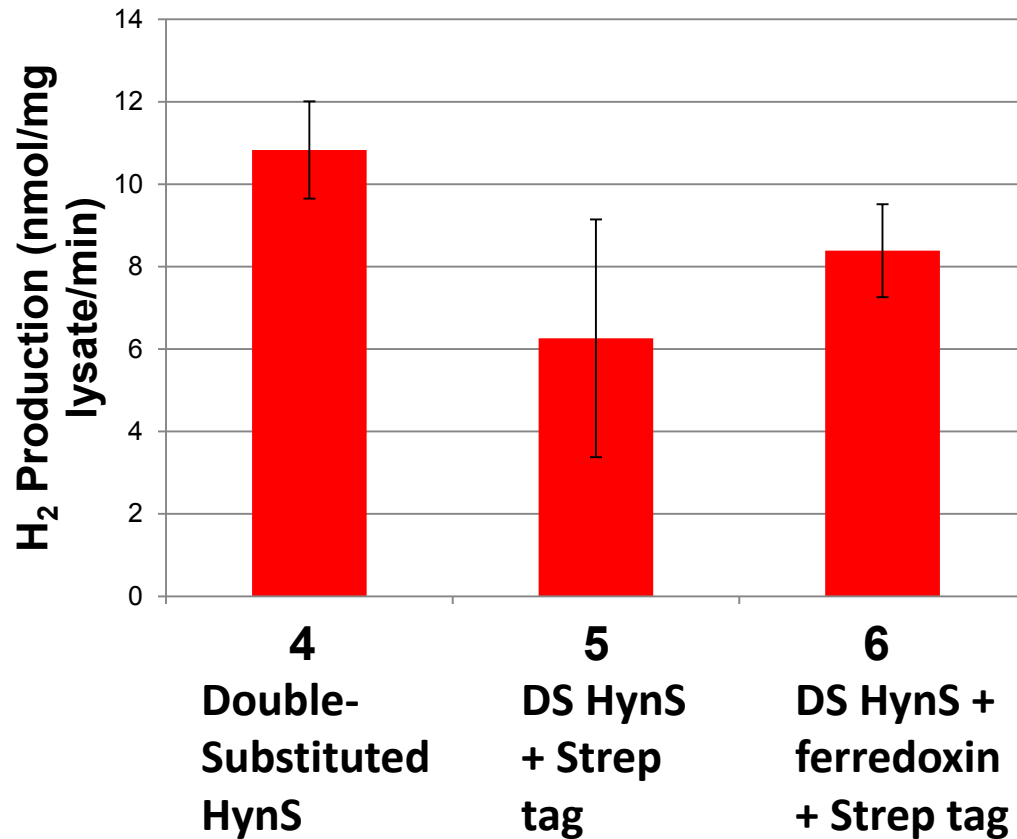
Blue = positive charge

**On track to complete Milestone “Improve hydrogenase-ferredoxin (Fd) electron transfer to enable 25-fold better Fd docking to the hydrogenase.” (11/13).**



# Task 2: Technical Accomplishments

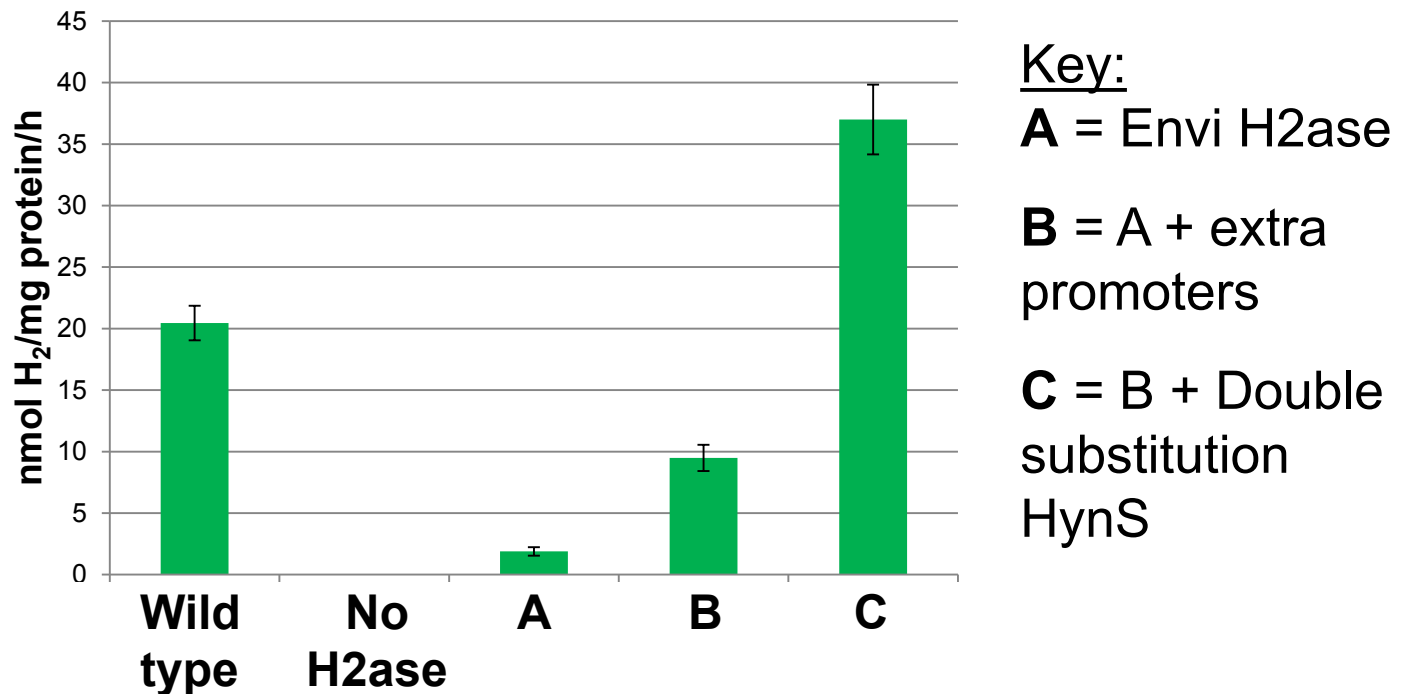
Ferredoxin-fusion H2ases are active when expressed in *E. coli*



**On track** to complete Milestone 2.5.3 “Construct an Fd-HynS fusion protein” (11/13).

# Task 2: Technical Accomplishments

## Activity improvements can be combined



**Exceeded Go-No Go Criteria** “Demonstrate 5x increase hydrogenase activity from environmental H2ase in cyanobacteria as measured by in vitro H<sub>2</sub> evolution assay.” (11/12)

# Collaborations

- NREL – Dr. Pin-Ching Maness
  - Expressing environmentally-derived hydrogenase in her *Synechocystis* sp. PCC 6803 system
  - Purified cyanobacterial ferredoxin
- Vanderbilt University – Dr. Carl H. Johnson and Dr. Yao Xu
  - Using circadian rhythm modification to enhance expression of O<sub>2</sub>-tolerant hydrogenases in cyanobacteria
  - Manuscript in preparation



# Proposed Future Work

## FY2013

- Continue optimization of promoter strength to achieve maximum expression of active hydrogenase.
- Continue to modify small subunit to increase ferredoxin binding.

## FY2014

- Combine all positive modifications into a single cyanobacterial strain and test for hydrogen production from light and water.

# Summary

- Developed strategies for increasing expression and activity of the environmentally-derived hydrogenase in cyanobacteria
  - Changed the frequency and strength of promoters.
  - Tested a novel T7 polymerase strategy for expression of hydrogenase.
  - Altered the FeS cluster ligation to increase H<sub>2</sub> evolution activity
- Developed strategies for increasing hydrogenase-ferredoxin interaction
  - Constructed a ferredoxin-hydrogenase fusion protein that maintains activity.